



## Review Article

# Metformin, Microbiota and Health



Graziele Silva Paz and Janaina Fernandes\*

*Nucleus of Multidisciplinary Research in Biology (Numpex-Bio), Federal University of Rio de Janeiro, Rio de Janeiro, Brazil*

Received: March 29, 2022 | Revised: July 12, 2022 | Accepted: August 11, 2022 | Published online: October 25, 2022

### Abstract

The intestinal microbiota is considered a large organ in the human body performing functions in the host that range from supporting digestion and absorption of nutrients from the diet to regulating the various processes in the host. Maintaining a diverse and stable microbiota is critical to maintaining host homeostasis and health. Studies have suggested the relationship between the microbial changes and the development of several pathologies. In this context, metformin, has shown to be a promising drug for the regulation of the microbiota, thus favoring the prevention and treatment of type 2 diabetes mellitus (T2DM), obesity, cancer, the inflammatory state of human immunodeficiency virus (HIV), heart disease, Alzheimer's disease and aging, and pathologies associated with dysbiosis. In this review, the main aspects on the importance of metformin's action on dysbiosis, and the factors that regulate the metformin uptake and activity as genetic polymorphisms and GLP-1 receptor activation were discussed.

### Microbiota and its role in the human body

Microbiota, whose composition and activity are associated with health and the development of diseases,<sup>1</sup> are composed of different microorganisms, including bacteria, bacteriophages, eukaryotic viruses, and fungi that inhabit body surfaces and cavities usually in a beneficial relationship for both parties.<sup>2</sup> Most microbiota's microorganisms are found in the gastrointestinal tract (mainly colon), and their composition are influenced by the type of birth, lifestyle, diet in the first years of life, medications, and host genetics.<sup>2</sup>

Human microbiota composition comprises approximately 1,000 species, where bacteria are considered the main constituents corresponding to the five main phyla: Bacteroidetes, Firmicutes, Actinobacteria, Proteobacteria, and Verrucomicrobia.<sup>3</sup> Studies have pointed out that the number of these microbial cells is around  $1 \times 10^{14}$  against  $1 \times 10^{13}$  of eukaryotic cells, thus indicating that the number of microbial cells is 10 times greater than the number of human cells.<sup>4</sup> Hence, the microbiota can be considered as a metabolic organ supporting important functions

in the host, such as digestion and absorption of food nutrients, the regulation of the immune system, regulation of the endocrine and neurological functions, alteration of the mechanism of the action of drugs, removing toxins, and the production of various compounds, including vitamins and short chain fatty acids (SCFAs).<sup>2,5</sup> The maintenance of diverse microbiota with a functionally stable microbiome contributes to the homeostasis and health of the host.<sup>6</sup>

Modulations in the composition of the microbiota, compromising the symbiotic relationship between the host and microorganisms in a process called dysbiosis, are often associated with the development of autoimmune diseases, obesity, diabetes, cancers, neurological diseases among others.<sup>7,8</sup> Dysbiosis can be classified into three types: (1) loss of beneficial microorganisms, (2) an increase in opportunistic pathogens, and (3) loss of microbiota diversity; despite this categorization, the three scenarios usually occur simultaneously.<sup>9</sup> Thus, factors capable of promoting changes in the composition of the microbiota are considered risk factors for diseases associated with a microbial imbalance.<sup>9</sup> Therefore, drugs able to restore the microbiota balance have become promising candidates for the treatment of these pathologies. In this context, metformin is an important candidate, so are several factors that regulate its uptake and activity.

### Metformin: Brief history and the main mechanisms of action

Metformin (1,1-dimethylbiguanide) is a drug of the biguanide class derived from guanidine, the active substance in Galega officinalis. The processes for its synthesis began in 1920, from the discovery of the antidiabetic properties in 1918 of guanidine.<sup>10</sup> Due to its high antihyperglycemic efficiency, without causing

**Keywords:** Gut microbiota; Dysbiosis; Metformin; GLP-1; Polymorphism.

**Abbreviations:** AD, Alzheimer's disease; AMPK, AMP-activated protein kinase; BCAAs, branched-chain amino acids; CRC, colorectal cancer; CVD, cardiovascular diseases; FMT, fecal microbiota transplantation; GLP1, glucagon-like peptide; HIV, human immunodeficiency virus; LPS, lipopolysaccharide; SCFAs, short chain fatty acids; T2DM, type 2 diabetes mellitus.

\***Correspondence to:** Janaina Fernandes, Nucleus of Multidisciplinary Research in Biology (Numpex-Bio), Federal University of Rio de Janeiro, Rio de Janeiro 21949-900, Brazil. ORCID: <https://orcid.org/0000-0002-3442-6101>. Tel: +55 21-99717-8702, E-mail: [janainaf@xerem.ufrj.br](mailto:janainaf@xerem.ufrj.br)

**How to cite this article:** Paz GS, Fernandes J. Metformin, Microbiota and Health. *J Explor Res Pharmacol* 2023;8(1):36–56. doi: 10.14218/JERP.2022.00032.

hypoglycemia and with a low risk of developing lactic acidosis, metformin has been used for about a century, and for some years, it has been considered the therapy of choice for type 2 diabetes mellitus (T2DM).<sup>10,11</sup>

In recent years, the action of metformin has been observed on other diseases like various types of cancer, cardiovascular and neurological diseases, obesity, among others.<sup>12</sup> Metformin's mechanism of action on adenosine monophosphate (AMP)-activated protein kinase (AMPK) is central to most diseases in which metformin acts<sup>13–15</sup> and involves the inhibition of complex I (CI) of the mitochondrial electron transport chain (ETC), consequently leading to mitochondrial dysfunction and energy impairment. Metformin's entrance into the mitochondrial matrix has led to a reduction in the adenosine triphosphate (ATP) levels and an increase in the adenosine diphosphate (ADP) and AMP levels, thus promoting the activation of AMPK.<sup>16</sup> AMPK works as an intracellular energy sensor that restores the energy homeostasis disrupted by metformin, which increases the catabolic processes and reduces the anabolic processes. Thus, processes such as protein, lipid, and nucleic acid synthesis end up being reduced compromising the progression of diseases dependent on these processes.<sup>17</sup>

### Metformin, microbiota modulation, and T2DM

T2DM is considered a metabolic syndrome characterized by impaired glucose metabolism resulting from a combination of genetic, environmental, and behavioral factors.<sup>18,19</sup> Epidemiological data for 2017 showed 462 million people worldwide diagnosed with T2DM, or corresponding to 6.28% of the world population.<sup>20</sup> This pathology is characterized by hyperglycemia associated with the scenario of insulin resistance leading to hyperinsulinemia, and in some cases, it could also be associated with reduced insulin production by pancreatic beta cells ( $\beta$ -cells) given the functional impairment.<sup>21</sup>

Some studies have pointed to differences in the microbiota between patients with T2DM and healthy patients,<sup>22,23</sup> which has been characterized by a reduction in the butyrate-producing strains (*Faecalibacterium prausnitzii* and *Roseburia intestinalis*) and an increase in the opportunistic pathogens (*E.coli* and *Clostridium ramosum*).<sup>18,24</sup> From this observation and several others, it was suggested that the involvement of the microbiota in the development and progression of T2DM passed through its metabolites. The involvement of the microbiota with T2DM was due to (1) compromising the integrity of the intestinal barrier, (2) reduced production of SCFAs, (3) the metabolization of bile acids, and (4) increased production of branched-chain amino acids (BCAAs)<sup>18,25,26</sup> (Fig. 1).

Metformin is orally administered and its concentration in the jejunum is estimated to be 30 to 300 times greater than in plasma indicating that the intestine is one of the main targets of metformin, in addition to constituting an important reservoir of this drug.<sup>27</sup>

This high uptake of metformin by the gastrointestinal tract influences the microbial profiles of T2DM patients treated with metformin compared to those who are untreated,<sup>28</sup> consequently revealing the impact of metformin in dysbiosis. At present, several studies have shown the ability of metformin to restore the present microbial profile before the development of the disease.<sup>18,29,30</sup> Microbiota regulation by metformin may also be induced by increasing or decreasing a particular strain.<sup>31</sup> Although changes in the microbiota are specific for each pathology in which metformin acts, in general, it is associated with an increase in various strains: *Verrucomicrobiaceae*, *Porphyromonadaceae*, *Rikenellaceae*, *Ak-*

*kermansia muciniphila*, *Prevotellaceae*, *Escherichia*, and *Shigella* sp., and reduction of the strains: *Lachnospiraceae*, *Rhodobacteraceae*, *Peptostreptococcaceae*, and *Clostridiaceae*.<sup>32</sup>

SCFAs produced by certain bacterial strains are involved in increasing insulin sensitivity and intestinal barrier integrity, in addition to mediating the activation of intestinal gluconeogenesis (mainly butyrate and propionate), thereby ensuring energy homeostasis and better regulation of glucose metabolism.<sup>33–35</sup> Reduction of strains that produce SCFAs is associated with T2DM, while its increase was observed in metformin-treated T2DM patients.

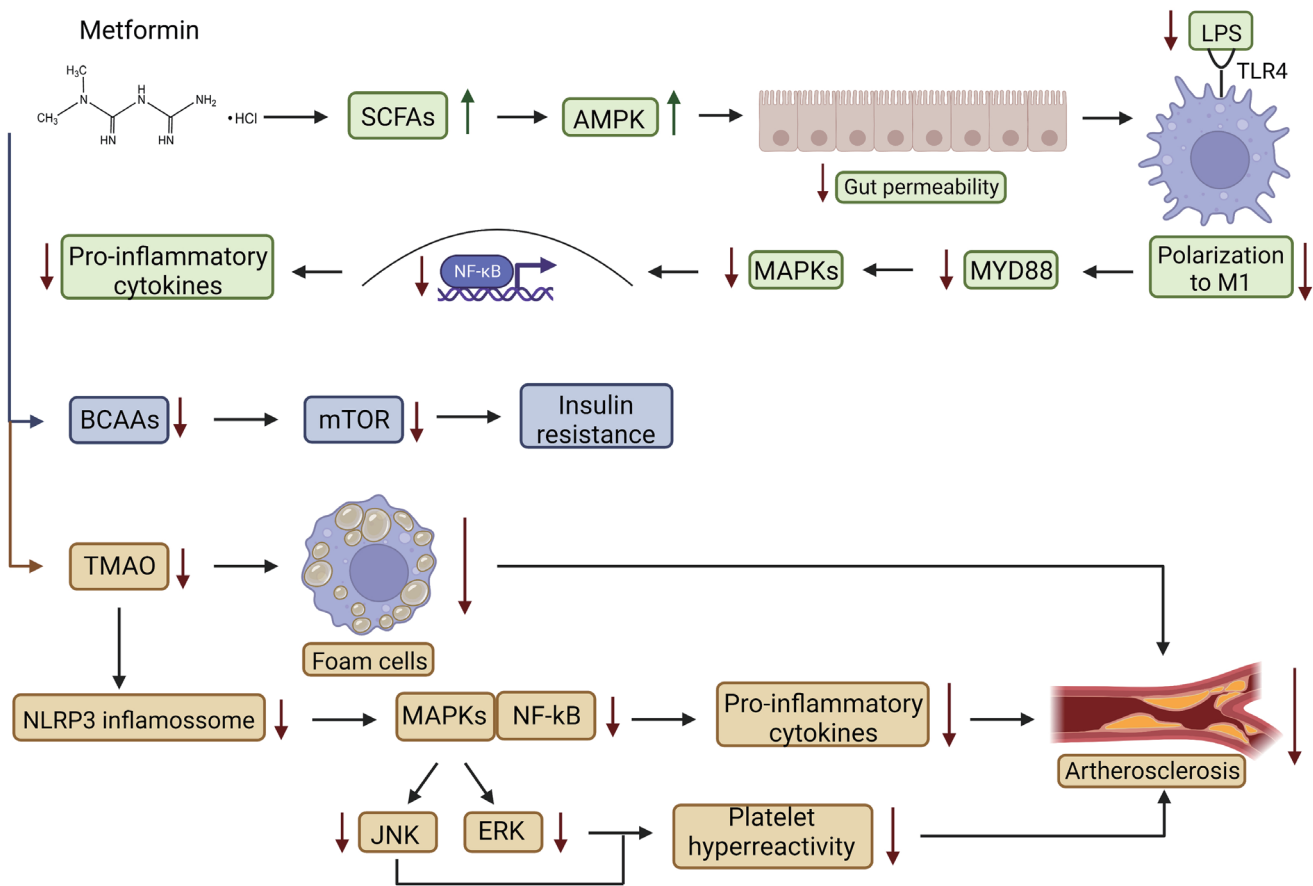
Microbial profile alterations induced by metformin in T2DM treated patients showed (1) an increase in the Firmicutes phylum, (2) an increase in the *Roseburia*, *Butyrivibrio*, and *Bifidobacterium* genera, (3) an increase of the strain *Akkermansia muciniphila*, and (4) a reduction of the *Bacteroides fragilis* strain.<sup>7</sup>

By modulating the microbiota, metformin was able to (1) preserve the gastrointestinal barrier by increasing the *Akkermansia muciniphila* strain and reducing Lipopolysaccharide (LPS)-producing strains, such as *Bacteroides fragilis*, therefore enabling the production of mucus in the gastrointestinal barrier that guaranteed its integrity and reduced intestinal permeability. It also prevented the release of LPS and the subsequent activation of toll-like receptor 4 (TLR4) on the surface of the macrophages leading to an inflammatory cascade that involved the activation of JNK and IKK $\beta$  and phosphorylation of IRS-1/2, which are considered as markers of insulin resistance and damage the glucose metabolism, respectively.<sup>5,30</sup> (2) This increased the synthesis of the SCFAs mainly through the increase of the *Akkermansia* and *Butyrivibrio* strains by increasing the intestinal integrity through the activation of AMPK which in turn regulated tight junctions between the enterocytes, thus reducing inflammation and improving the metabolism of glucose and lipids.<sup>18</sup> (3) Metformin regulated the bile acids, associated with the reduction of the *Bacteroides fragilis* strain that allowed an increase in the level of glycochenodeoxycholic bile acid (GUDCA), which plays an anti-inflammatory role as a farnesoid X receptor (FXR) antagonist leading to an increase in insulin sensitivity and maintenance of glucose homeostasis.<sup>36</sup> (4) It reduced the levels of the BCAAs (leucine, isoleucine, and valine), which were considered as risk factors for T2DM due to their action on the mammalian target of rapamycin (mTOR) pathway to induce insulin resistance (Fig. 1).<sup>7,37–39</sup>

Even though the use of metformin became widespread, in recent years, the development of glucagon-like peptide (GLP)-1 receptor agonists came to aid in controlling T2DM. GLP-1 is a 30-amino-acid peptide hormone produced in intestinal epithelial endocrine L-cells through the processing of proglucagon. Along with insulin secretion and a sensitivity increase, GLP-1 reduces glucagon secretion and improves hepatic glucose metabolism,<sup>40</sup> therefore leading to a reduction in body weight, inflammation, and altered bile acid composition resulting in an improvement of the patient's health.

Moreover, it was suggested that treatment with GLP-1 receptor agonists could influence the composition of microbiome,<sup>41</sup> and target both T2DM and obesity simultaneously.<sup>41,42</sup> GLP-1 peptide was augmented for its microbial diversity in hyperglycemic and obese mice, but it was unclear if this effect was due to the treatment with the peptide or a secondary effect of weight loss.

Associated with the modulation of gut microbiota by metformin, a study by Koh *et al.* showed that the microbial metabolite imidazole propionate, generally found in high levels among T2DM patients and related to glucose intolerance,<sup>43</sup> impaired the effect of metformin on the glycemic reduction in diabetic mice inhib-



**Fig. 1. Metformin action against T2DM, obesity, CRC, HIV inflammation, and CVDs through microbiota and molecular alterations.** Through microbiota modulation, metformin enhanced the SCFAs strains, leading to AMPK activation and the reduction of gut permeability. This resulted in decreased LPS release in circulation and its recognition by TLR4 in macrophages. Furthermore, it prevented the polarization of macrophages to the M1 phenotype, the recruitment of protein adaptors (such as MYD88), and the activation of MAPKs and NF-κB, thereby reducing the secretion of pro-inflammatory cytokines. Beyond its action against inflammation in T2DM and CVDs, metformin reduced the BCAAs levels in these patients by avoiding mTOR activation and insulin resistance, a risk factor for T2DM and CVDs. Due to the importance of the TMAO levels for the development of CVDs, metformin reduced TMAO through microbiota modulation by avoiding macrophage transformation into foam cells (a hallmark of AS) and the activation of NLRP3 inflammasome, consequently reducing MAPKs (JNK and ERK), NF-κB activation, platelet hyperreactivity, the secretion of pro-inflammatory cytokines, and the risk of AS development. Green squares: metformin action in T2DM, obesity, CRC, HIV inflammation and CVDs; blue squares: metformin action in T2DM and CVDs; orange squares: metformin action in the CVDs. AMPK, AMP-activated protein kinase; AS, atherosclerosis; BCAAs, branched chain amino acids; CRC, colorectal cancer; CVDs, cardiovascular diseases; ERK, extracellular signal-regulated kinase; JNK, c-Jun N-terminal kinase; LPS, lipopolysaccharide; MAPKs, mitogen-activated protein kinase; mTOR, mammalian target of rapamycin; MYD88, myeloid differentiation protein 88; NF-κB, nuclear factor kappa B; NLRP3, NOD-LRR- and pyrin domain-containing protein 3; SCFAs, short chain fatty acid; T2DM, type 2 diabetes mellitus; TLR4, toll-like receptor 4; TMAO, trimethylamine N-oxide. Created with www.biorender.com.

iting the phosphorylation of the AMPK active site T172 through the activation of p38 gamma, but the effects of metformin were recovered after the inhibition of p38 gamma. This indicated that a microbial metabolite could be associated with metformin response variations among T2DM patients.

Understanding the importance of the regulation of microbiota exerted by metformin, so that it could exert its mechanisms of action, a study by Pryor *et al.* evaluated the relationship between nutrients, microbiota, and metformin action. In this study, it was demonstrated that the nutrients from the amino sugars, peptides, amino acids, and nucleotides classes would be important to the regulation of dysbiosis found in diseases, such as T2DM, by metformin. These nutrients would favor the growth of strains like *Escherichia coli*, where metformin would promote the enhancement of galactose and the phosphotransferase system (PTS) of the transcription factors Crp and ArgR, thereby indicating the importance

of the bacterial PTS-Crp to the metformin action. The PTS is a system associated with the uptake of carbohydrates and downstream regulation of Crp; as such, possibly the Crp with its cofactor AMP could control the transcription of different genes that could alter the metabolism and be associated with an extended lifespan, including the enhancement of the agmatine production by the bacteria that would exert an important role on metformin action in T2DM.<sup>44</sup>

The glycoregulation action of metformin could also be explained by its effect on the enhancement of the expression of the sodium glucose cotransporter-1 (SGLT1) in the upper small intestine that was demonstrated in rodents<sup>45</sup> and humans.<sup>46</sup> This transporter acts on the uptake of glucose and the secretion of GLP-1.<sup>47</sup> As a way to elucidate how metformin promotes the expression of SGLT1, Bauer *et al.* showed that metformin induced changes in the microbiota of the upper small intestine by enhancing strains of *Lactobacillus* genus

that acted on the enhancement of SGLT1, an important transporter for the anti-hyperglycemic effect of metformin.<sup>48</sup>

### **Metformin, microbiota, and gestational diabetes mellitus (GDM)**

Gestational diabetes mellitus (GDM) is a common metabolic complication among pregnant women, which is characterized by hyperglycemia that brings risks to the mother and baby.<sup>49</sup> The treatment involves changes in the lifestyle to the use of insulin and metformin, in which the latter has gained more preference due to less side effects.<sup>50,51</sup> Even though it was already demonstrated that metformin altered the microbiota, there have been a number of studies on the microbiota alteration in women with GDM under metformin use. In this context, the study of Molina-Vega *et al.* showed that women treated with metformin had lower blood glucose and weight gain, and these effects would be associated with the enhancement of the Proteobacteria, Enterobacteriaceae, and Coprococcus strains involved with a lower body mass index (BMI) and blood glucose, and with the decrease of the Firmicutes and Peptostreptococcaceae, strains associated with a higher BMI and blood glucose. Therefore, metformin promoted changes in the microbiota of women with GDM as a way to corroborate its effects in GDM.<sup>52</sup>

### **The impact of probiotics, prebiotics, and metformin in a microbiome**

The use of probiotics (live microorganisms administered to the host at a certain concentration) and prebiotics (non-digestible nutritional ingredients that stimulate the growth of strains) beneficial to the host in combination with metformin could promote valuable changes in the composition of the intestinal microbiota<sup>53</sup> by corroborating an antihyperglycemic effect and improving the response to metformin in intolerant T2DM patients.<sup>54,55</sup> The combination of metformin and probiotics (*Lactobacillus plantarum*, *Lactobacillus bulgaricus*, *Lactobacillus gasseri*, *Bifidobacterium breve*, *Bifidobacterium animalis* sbsp. *lactis*; *Bifidobacterium bifidum*, *Streptococcus thermophilus* and *Saccharomyces boulardii*)<sup>55</sup> increased the SCFA-producing strain *Anaerotruncus colihominis* and the metabolic pathways associated with butyrate production (SCFA) in patients treated with this combination, which was not observed with treatments using only one or the other. In addition, associated with the increase in the SCFAs in patients treated with the combination, an improvement in blood glucose and attenuation of insulin resistance was observed. Hence, this indicated that the use of probiotics could increase the antihyperglycemic efficiency of metformin in T2DM patients through microbiota modulation.<sup>55</sup>

Prebiotics could also aid the effects of metformin. In Shin *et al.*'s clinical study with T2DM patients, the subjects were subjected to a placebo or *Scutellaria baicalensis* (SB) root (antidiabetic action) with metformin for a period of eight weeks, and its effects on the pro-inflammatory markers (TNF- $\alpha$  and IL-6) and on the antihyperglycemic action of metformin were evaluated. A reduction in glucose intolerance associated with changes in the intestinal microbiota was observed in patients treated with SB+metformin with an increase in the *Lactobacillus* and *Akkermansia* strains,<sup>53</sup> which was considered important for glycemic and insulin homeostasis.<sup>56,57</sup> Therefore, SB acted as a prebiotic and improved the antidiabetic efficiency of metformin by modulating the intestinal microbiota.<sup>53</sup>

It is also important to emphasize that because T2DM is a risk factor for obesity, cardiovascular diseases (CVDs) and cancer, in addition to compromising healthy aging, the use of probiotics and

prebiotics as adjuvants to metformin in T2DM could reduce the risk of developing these diseases.

### **Metformin and obesity**

Obesity is a multifactorial disease whose incidence has tripled since 1975, and responsible for affecting more than 650 million adults, and 340 million children and adolescents according to data from the World Health Organization (WHO) in 2016.<sup>58</sup> Furthermore, the pre-obesity stage affected more than 1.9 billion adults according to the WHO data in 2016.<sup>58</sup> In Brazil, between 2016 and 2017, obesity increased by 60% among people over 18 years of age.<sup>59</sup> Overweight and obesity are defined as the high accumulation of fat that causes important impacts on health and is related to the development of several diseases, such as arterial hypertension, dyslipidemia, CVDs, T2DM, and cancer.<sup>60</sup>

Even though some genetic and epigenetic factors can contribute to the development of obesity, the interaction between genetic and environmental factors, such as diet and lifestyle leads to the imbalance between energy obtained and spent.<sup>61</sup>

In addition, many studies suggest the participation of the intestinal microbiota in the development of obesity.<sup>62,63</sup> The first study that demonstrated the association between dysbiosis and obesity was carried out by Ley *et al.*, where a 50% reduction in the genus *Bacteroidetes* and 50% increase in the genus *Firmicutes* in obese animals was observed through metagenomic analysis; moreover, these animals showed an increase in the methanogenic Archaea strains, such as *Methanobrevibacter smithii*, which in turn were associated with a higher efficiency of fermentation.<sup>64</sup> The study was extended to humans showing a similar profile.<sup>65</sup> Other studies also observed an increase in the *Firmicutes/Bacteroidetes* ratio in obese people.<sup>66–69</sup> However, some others found a contrary profile or did not detect changes between these strains.<sup>70–73</sup>

Other strains were also altered in obesity. Some studies showed a lower abundance of the genus *Bifidobacterium* in obese individuals.<sup>68,71,74</sup> However, only a few species belonging to this genus had their anti-obesity action confirmed in animals,<sup>75</sup> which was associated with their anti-inflammatory effect. Additionally, opportunistic pathogens were often associated with obesity like *Fusobacterium*, *Escherichia/Shigella*, *Staphylococcus aureus* that were observed in pregnant women and overweight children, and *Enterobacteriaceae* was observed in pregnant and overweight women.<sup>71,68–77</sup> Unlike opportunistic pathogens, beneficial strains were generally present in low concentration in obese individuals, such as *Akkermansia muciniphila*.<sup>78</sup> Hence, the development of obesity is mainly associated with its involvement with the reduction of SCFAs, inflammatory processes, and energy regulation.<sup>79–81</sup>

The intestinal microbiota are also able to ferment polysaccharides present in the diet that are not digested by humans, thus generating SCFAs.<sup>80</sup> Once absorbed, SCFAs could be transferred to hepatocytes, where they would mediate lipogenesis and gluconeogenesis as substrates.<sup>82,83</sup> Additionally, they would act as signaling molecules binding to free fatty acid receptors 2 (GPR43 or FFAR2) and 3 (FFAR3 or GPR41), and would be expressed mainly by intestinal epithelial cells.<sup>84</sup> Through these receptors, SCFAs would promote greater energy uptake from food and fat accumulation leading to an increase in body mass.<sup>79</sup> SCFAs also promote the inhibition of the fasting-induced adipocyte factor (FIAF), which inhibits lipoprotein lipase (LPL) activity resulting in the accumulation of triglycerides in adipocytes.<sup>80</sup>

Furthermore, the inflammatory process is an important contributor to obesity, and some studies have already demonstrated the

presence of pro-inflammatory cytokines during the progression of obesity.<sup>85,86</sup> The relationship between microbiota, obesity, and inflammation was found to be associated with LPS, which is related to the deposition of adipose tissue, the inflammatory process, and insulin resistance.<sup>87,88</sup>

The microbiota of obese individuals is also considered a little diverse and contributes in different ways to the development and progression of obesity.<sup>89</sup> Thus, drugs capable of regulating the microbiota and circumventing obesity-associated dysbiosis are promising candidates for the treatment of obesity.<sup>18</sup>

Likewise, several studies have reported weight loss promoted by metformin both in diabetic patients<sup>90,91</sup> and in non-diabetic patients.<sup>92,93</sup> Metformin plays its anti-obesity effects through 1) increased expression of the fibroblast growth factor 21 (FGF21), which is associated with increased lipolysis,<sup>94</sup> 2) increased activity of brown adipose tissue (BAT), a tissue rich in mitochondria capable of increasing thermogenesis,<sup>95</sup> and 3) reduced appetite associated with lactate production<sup>96</sup> and increased levels of GLP-1, a molecule that acts in the regulation of glucose metabolism.<sup>97</sup>

The activation of AMPK by metformin plays an important role in its actions, as it is associated with the reduction of blood glucose, insulinemia, cholesterol synthesis, and lipogenesis, thereby allowing metformin to perform its anti-obesity and antidiabetic effects.<sup>98–102</sup> AMPK is expressed in different tissues; however, little is known about the role of intestinal AMPK in the regulation of energetic and glucose homeostasis. In this context, a study by Zhang *et al.* created a knockout mice model for intestinal AMPK and observed its importance for metformin's action. The knockout group treated with metformin showed a reduction in the effects of this drug on (1) weight loss, (2) the improvement of glucose tolerance, (3) the accumulation of lipids in the liver, and (4) the reduction of the triglycerides, cholesterol, and insulin levels. These observations indicated the importance of intestinal AMPK for the metformin action in obesity, diabetes, and associated diseases. Throughout the study, the inhibition of intestinal AMPK was observed for its association with the impairment of microbiota alterations that would be important for metformin action.<sup>103</sup>

Along with its molecular action, metformin could exert its anti-obesity effects through the modulation of the microbiota. Studies have also shown that metformin promoted the increase of the *Akkermansia muciniphila* strain usually found in low concentrations in overweight people, thus performing an anti-obesity action.<sup>56,104</sup> In addition, metformin was able to reduce the level of *Escherichia coli* usually associated with being overweight.<sup>105</sup> Probiotic strains are reported to have beneficial effects for weight loss; in this context, metformin has been shown to increase the beneficial bacteria *Prevotella* and *Lactobacillus* strains associated with weight loss.<sup>104,106</sup> In obese people, there is a large production of SCFAs; however, this high production is contrary to the diversity of SCFA-producing strains in the microbiota of these individuals.<sup>89</sup> In this context, metformin was able to increase the diversity of SCFA-producing bacteria, such as *Allobaculum*, *Bacteroides*, *Blautia*, *Butyrivococcus*, and *Phascolarctobacterium*.<sup>106</sup> By increasing the diversity of the producers, the probability of SCFA absorption and performance of beneficial effects would be increased, such as increased intestinal integrity, preventing the release of inflammatory molecules, and triggering anti-obesity effects.<sup>89</sup>

Associated to microbiota modulation to exert its anti-obesity effect, some studies demonstrated that metformin reduced the LPS-associated inflammation through the downregulation of TLR4, a key regulator of this process. Other studies indicated that through AMPK activation, metformin promoted the macrophage polariza-

tion to M2 phenotype triggering anti-inflammatory effects (Fig. 1).<sup>107</sup>

Therefore, metformin may have anti-obesity effects either through its molecular mechanisms of action or through the modulation of the microbiota.

### Metformin and colorectal cancer (CRC)

Data from GLOBOCAN in 2020 pointed to colorectal cancer (CRC) as being responsible for 1.9 million cases and 935,000 deaths, thereby ranking third among the 10 most common types of cancer and second among those that most lead to death worldwide.<sup>108</sup> Advanced age and male individuals were highly associated with the risk of developing CRC.<sup>109</sup> Both genetic and environmental factors were also among the risk factors for CRC among which environmental factors could be highlighted: smoking,<sup>110</sup> alcoholism,<sup>111</sup> obesity,<sup>112</sup> the consumption of red and processed meats,<sup>113</sup> T2DM,<sup>109</sup> and microbiota with emphasis on the *Fusobacterium nucleatum*<sup>114</sup> and *Bacteroides fragilis*<sup>115</sup> strains.

The development of CRC begins with the formation of an aberrant crypt that evolves into a neoplastic lesion (polyp) providing progression to CRC, whose process takes around 10–15 years.<sup>109</sup> Surgical removal of polyps is usually indicated to reduce the risk of progression to CRC,<sup>116</sup> however, the chance of new polyps and of CRC development is high.<sup>117</sup> In this context, some chemotherapy drugs have been used as the prophylaxis for the development of CRC, such as non-steroidal anti-inflammatory drugs; however, the risk of developing cardiovascular diseases as a side effect is high.<sup>118</sup> This scenario has led to the search for safer drugs to exercise this chemoprophylaxis and even treatment for CRC with metformin as a strong candidate.<sup>119</sup>

The use of metformin in cancer is mainly associated with the inhibition of CI of the ETC resulting in energy depletion (increased AMP/ATP and ADP/ATP ratio) and activation of the energy sensor AMPK limiting tumor proliferation.<sup>120</sup> Several studies have already reported the antitumor effect of metformin on CRC of being able to reduce or inhibit the formation of aberrant crypt foci (ACFs) and colorectal polyps in *in vivo* studies,<sup>121</sup> clinical,<sup>122</sup> and *in vitro*.<sup>123</sup> In addition to its molecular antitumor effects, metformin plays an anti-CRC action through its ability to regulate the microbiota.<sup>29</sup>

CRC patients have microbiota with low diversity compared to healthy individuals.<sup>124</sup> However, it is not possible to standardize the characteristic microbiota of CRC.<sup>125</sup> Although the dysbiosis found in patients with CRC is mainly associated with bacterial strains, some fungi are also related to this neoplasm. Even though fungal diversity was not affected in patients with CRC, it was found to alter the Basidiomycota/Ascomycota ratio and the Malasseziomycetes class, in addition to the reduction of Saccharomycetes and Pneumocystidomycetes.<sup>126</sup>

One of the main bacterial strains associated with CRC is *Fusobacterium nucleatum*,<sup>127,128</sup> This strain is responsible for creating a tumor microenvironment for CRC through the TLR4/MYD88/NF- $\kappa$ B pathway by generating a pro-inflammatory environment and enabling epithelial-mesenchymal transition (EMT) through the activation of the nuclear factor kappa B (NF- $\kappa$ B), which is considered an important factor for CRC invasion and metastasis.<sup>129</sup> Thus, *F. nucleatum* is considered a biomarker for CRC, and its detection at high concentrations is related to a poor prognosis and low survival expectancy.<sup>130,131</sup> Additionally, *Enterococcus faecalis* is generally found at high levels in patients with CRC and is associated with oxidative stress through the generation of hydroxyl radicals

that act as mutagens, promote genetic instability, and increase the risk of developing CRC.<sup>132,133</sup>

Another important feature related to CRC is the high levels of *E. coli*.<sup>134</sup> Even though it is considered a commensal strain, *E. coli* produces the colibactin genotoxin inducing DNA damage and increasing the risk for CRC.<sup>85,135</sup> On the other hand, probiotic bacteria (SCFA producers), such as *Clostridium butyricum* (producing butyrate), are generally found in low concentrations or are non-existent in CRC patients.<sup>125</sup> Moreover, strains producing SCFAs, mainly butyrate, have anti-inflammatory, pro-apoptotic action and protect DNA against damage, which exerts beneficial effects in the fight against CRC.<sup>136,137</sup> Metformin interacts with *F. nucleatum* by reducing its levels and antagonizing the progression of CRC.<sup>138</sup> Beyond reducing the levels of this strain, metformin is associated with the inhibition of the TLR4/MYD88/NF-KB pathway,<sup>139</sup> thus reducing the inflammation and metastasis in CRC (Fig. 1). *F. nucleatum* is associated with chemotherapy resistance and post-chemotherapy recurrence in CRC.<sup>138</sup> Therefore, metformin, by reducing the *F. nucleatum* levels, could increase the chemotherapeutic efficiency and reduce the risk of recurrence.

Patients with CRC usually have low levels of *Akkermansia muciniphila* and *Bifidobacterium*,<sup>140,141</sup> a scenario that is reversed by metformin. *A. muciniphila* is a butyrate-producing strain associated with the antitumor effects of metformin, and mediates the anti-inflammatory, pro-apoptotic, and antitumor effects of this drug,<sup>142,143</sup> while *Bifidobacterium* is a probiotic strain performing antitumor effects through its pro-apoptotic action.<sup>144</sup>

Bile acid levels are generally elevated among CRC patients and are considered important for the carcinogenesis of this type of cancer.<sup>145,146</sup> Studies have shown the ability of metformin to reduce the abundance of bile acids producing strains, such as *Bacteroides fragilis*, which is found in high concentrations in CRC patients<sup>147</sup> antagonizing CRC carcinogenesis.<sup>36</sup>

Another strain associated with CRC is *Helicobacter pylori*, a bacterium that infects the mucosal layer above the gastric epithelium and causes gastritis that can progress to chronic stomach inflammation and in some cases lead to CRC.<sup>148</sup> Metformin is able to inhibit *H. pylori* by preventing this bacterial strain from triggering the development of CRC.<sup>149</sup>

Therefore, metformin antagonizes the development of CRC both by its molecular effects and by modulating the intestinal microbiota.

### Metformin and its action on HIV-associated inflammation

The human immunodeficiency virus (HIV) targets the host's immune system. In 2020, it was estimated that about 1.5 million people had been infected with HIV, thereby reaching a total of 37.7 million people infected worldwide.<sup>150</sup>

HIV targets CD4+ T cells of the intestinal mucosa by promoting the reduction of this cell type and compromising the intestinal epithelium, which allows the translocation of microbial products to the circulatory system and triggers a systemic inflammatory response.<sup>151</sup> By compromising the immunity of the intestinal epithelium, HIV triggers dysbiosis.<sup>152,153</sup> Antiretroviral therapy (ART) has also revolutionized HIV treatment and increased the life expectancy and quality of life of HIV-positive individuals. However, even regulating a viral load, CD4+ T cell concentration and reducing inflammation, ART cannot restore the immune response to the level of HIV-negative individuals.<sup>154</sup> Therefore, intestinal impairment, dysbiosis, and chronic inflammation remain in HIV-positive patients leading to the development of dyslipidemia, cardiovascu-

lar diseases, depression, and cancer.<sup>155–157</sup>

Furthermore, the gastrointestinal tract is essential for the HIV pathogenesis, as it is the main site of replication and viral reservoir<sup>158</sup> given the high expression of the CCR5 receptor on CD4+ T cells that functions as a co-receptor for HIV.<sup>159</sup> By compromising intestinal immunity, HIV triggers dysbiosis and systemic inflammation.<sup>151</sup> In HIV positive individuals, dysbiosis is characterized by an increase in Proteobacteria, Enterobacteria, and Fusobacteria, in addition to a reduction in Ruminococcaceae, Bacteroidia, Anaerovibrio, Bifidobacterium, and Clostridium.<sup>157,160,161</sup> HIV-associated dysbiosis is characterized by the reduction of strains related to the maintenance of intestinal integrity among which are *A. muciniphila* and the butyrate-producing strains, *Roseburia*, *Coprococcus*, *Faecalibacterium*, and *Eubacterium*.<sup>162–165</sup>

Likewise, metformin is able to benefit HIV-positive patients by reducing systemic inflammation through the (1) inhibition of NF-κB, (2) reduction of cytokine production, such as TNF-α and IL-1, and (3) through changes in the microbiota.<sup>33,36,104,166–168</sup> In this context, previous studies have demonstrated the increase of the *A. muciniphila* and butyrate-producing strains in HIV-positive patients by metformin, which has enabled an increase in intestinal integrity, consequently avoiding LPS release and associated inflammation.<sup>169,170</sup>

Thus, by regulating the microbiota in HIV-positive patients, metformin promotes an increase in intestinal integrity and a reduction in systemic inflammation, where it supports the ART treatment and reduces the risks of development of pathologies associated with inflammation.

### Metformin and cardiovascular disease (CVD)

CVDs consist of a group of pathologies associated with the heart and blood vessels, including coronary heart disease, cerebrovascular disease, peripheral arterial disease, rheumatic heart disease, congenital heart disease, pulmonary embolism, and deep vein thrombosis.<sup>171</sup> The WHO's data indicate that CVDs were responsible for 17.9 million deaths in 2019, which corresponded to 32% of global deaths.<sup>171</sup> Studies have further shown that the main risk factors for CVDs involve T2DM, obesity, hypertension, dyslipidemia, platelet hyperactivity, and atherosclerosis (AS).<sup>172–176</sup> While genetic factors are responsible for 20% of CVD cases,<sup>177</sup> a growing number of studies have pointed to the importance of the intestinal microbiota for CVDs.<sup>178,179</sup>

Additionally, Bacteroidetes and Firmicutes strains comprise more than 90% of the intestinal microbiota, and in this context, an increase in the Firmicutes/Bacteroidetes ratio was observed among patients with CVD,<sup>180,181</sup> which was associated with inflammation and diabetes mellitus, both considered risk factors for CVDs.<sup>5</sup> The same was observed in individuals with CVDs regarding Enterobacteriaceae and *Streptococcus* spp., in addition to a reduction in SCFA-producing strains, such as *Roseburia intestinalis*, *Faecalibacterium prausnitzii*, and *Eubacterium rectale*.<sup>182,183</sup> which could lead to the development of risk factors for CVDs, such as obesity, dyslipidemia, insulin resistance, and oxidative stress.<sup>184,185</sup> An increase in trimethylamine oxide (TMAO) producing strains, an important metabolite for the development of CVDs, was also observed.<sup>186–188</sup> TMAO is produced from the metabolism of phosphatidylcholine and choline by microbial enzymes that generates trimethylamine (TMA), which in turn is converted in the liver by flavin-containing monooxygenases, mainly by FMO3, resulting in TMAO.<sup>189,190</sup> Moreover, the increase in TMAO production is associated with increased inflammation and CVDs through the activa-

tion of SR-A and CD36 receptors on the macrophages that promote the recognition and uptake of oxidized low-density lipoprotein (ox-LDL), a risk factor for CVD associated with AS. The uptake of ox-LDL stimulates the transformation of the macrophages into foam cells, which is an early hallmark of AS. The high levels of TMAO also induce the activation of the NLRP3 inflammasome, a lipoprotein complex formed through the activation of pattern recognition receptors (PRRs) of great importance for AS that promote the activation of caspase 1 and the consequent secretion of pro-inflammatory cytokines IL-1 $\beta$  and IL-18 contributing to AS. TMAO is also associated with the activation of MAPKs, JNK, and ERK, which can lead to platelet hyperreactivity and to AS, in addition to promoting the activation of NF- $\kappa$ B that acts on the secretion of pro-inflammatory cytokines, such as IL-1 and TNF- $\alpha$  contributing to the development of AS (Fig. 1).<sup>191</sup>

Another microbial metabolite associated with CVDs are bile acids, especially secondary ones.<sup>192</sup> Primary bile acids are metabolized by bile salt hydrolase enzymes (BSHs) present in bacterial strains, such as in *Methanobrevibacter smithii*, *Clostridium*, and *Enterococcus*,<sup>193,194</sup> into secondary bile acids, such as deoxycholic acid (DCA) and lithocholic acid (LCA) which in turn are agonists of the TGR5 and FXR receptors.<sup>195,196</sup> By binding to their receptors, secondary bile acids promote macrophage activation, inflammation, and AS.<sup>197,198</sup>

Beyond SCFAs, TMAO, and bile acids, another microbial metabolite important to CVDs is the BCAAs that are associated to many risk factors for the development of CVDs, such as obesity, insulin resistance, arterial hypertension, dyslipidemia, and coronary disease indicators.<sup>37,199–202</sup> In this context, Tobias *et al.* demonstrated the relationship between the high levels of BCAAs in plasma and CVDs, which was probably due to the impairment of the BCAAs catabolism leading to its accumulation and enhancing the risk of AS associated with insulin resistance.<sup>203</sup>

Metformin acts on cardioprotection due to its antidiabetic, anti-obesity and anti-dyslipidemia mechanisms of action.<sup>96,204,205</sup> Furthermore, the cardioprotective effects of metformin may be mediated by its action on the regulation of the intestinal microbiota.<sup>206</sup> *Akkermansia muciniphila* is a strain associated with health and good cardiac parameters,<sup>78</sup> which increased concentration has been linked to metformin, hence preventing pro-inflammatory conditions and reducing the risk of developing CVDs.<sup>56,207–209</sup> In addition, the role of metformin in the production of bile acids exerts its cardioprotective effects since metformin is associated with a reduction in the BSH activity of the *Bacteroides fragilis* strain and the lower content of Firmicutes, such as *Clostridium perfringens*, reducing inflammation and the progression of AS.<sup>210–213</sup>

Metformin is able to increase the concentration of the SCFAs producing strains of *Butyrivibrio*, *Bifidobacterium bifidum*, and *Megasphaera* contributing to an anti-inflammatory effect and cardioprotection.<sup>167,56</sup> Studies have indicated that metformin could reduce the levels of TMAO by regulating the producing microbiota as a way to exert its cardioprotective effects.<sup>214,215</sup> In the study by Su *et al.*, metformin was able to reduce the TMAO levels through increasing the *Akkermansia* and *Bifidobacterium* strains (negatively related to TMAO) and reducing the *Lachnospirillum* and *Ruminiclostridium* strains (positively related to TMAO) *in vivo* (Fig. 1).<sup>215</sup> Furthermore, metformin was able to reduce the levels of TMA and TMAO through its action on TMAO metabolism<sup>209</sup> leading to a reduction of the inflammatory cascade associated to TMAO.

Associated with the metformin action in the BCAAs, thus inducing its reduction in T2DM patients through the modulation of

the microbiota and its action as an mTOR antagonist, metformin reduces insulin resistance and the contribution of this risk factor for the development of CVDs.<sup>7,38–40</sup>

### Metformin and cognitive function: Therapeutic contribution to Alzheimer's disease

Dementia is a pathology associated with impaired cognitive function and aging. It is estimated that 55 million people suffer from dementia worldwide with an annual incidence of about 10 million new cases per year. It is considered the seventh most fatal disease worldwide; additionally,<sup>216</sup> AD is responsible for 60–80% of diagnosed cases of dementia.<sup>217</sup>

AD is clinically characterized by (1) cognitive and memory impairment, (2) the onset of psychiatric symptoms and behavioral problems, and (3) impairment of daily tasks.<sup>218</sup> The initial symptomatology of AD involves impairment of recent memories, changes in unconscious behavior, as well as changes in language and speech.<sup>219</sup> At the molecular, biochemical and cellular level, AD is characterized by (1) cell death, (2) impairment of energy metabolism, (3) hyperactivation of signaling pathways, (4) deposition of amyloid beta proteins (A $\beta$ ), (5) mitochondrial impairment, (6) oxidative stress, and (7) DNA damage.<sup>220–223</sup>

Despite the various cellular, biochemical, and molecular factors related to AD, the deposition of A $\beta$  is considered its main feature.<sup>219</sup> The A $\beta$  deposition promotes the hyperphosphorylation of the tau protein (p-tau), neuroinflammation, oxidative stress, and neuronal degeneration.<sup>224</sup> Studies have indicated that the accumulation of A $\beta$  proteins is considered the initiating factor of AD and is important for the formation of the tau aggregates, in addition to promoting the activation of microglia cells and astrocytic recruitment culminating in local inflammation and providing neuronal impairment and death.<sup>219–221,225</sup> This neurodegenerative disease can be caused by both hereditary and non-hereditary factors, such as aging, lifestyle, and environmental factors.<sup>220</sup> Among the environmental factors, the intestinal microbiota stands out.<sup>226</sup>

Studies have already demonstrated that individuals with changes in the microbiota are more likely to develop neurological diseases, such as AD.<sup>227,228</sup> Based on this observation, other studies found the ability of the microbiota to regulate the behavior and brain function contributing to the development of neurological diseases due to the microbiota-gut-brain axis.<sup>229–234</sup> In this context, the study by Morris *et al.* showed that 85% of individuals with dementia had a different microbiota from healthy individuals,<sup>235</sup> hence indicating the importance of dysbiosis for the development of neurological diseases, such as AD. Table 1 highlights the main studies related to dysbiosis with AD.<sup>228,235–237</sup>

A clinical study performed on AD patients showed that the *Bacillus subtilis*, *E. coli*, *Klebsiella pneumoniae*, *Mycobacterium* spp., *Salmonella* spp., *Staphylococcus aureus*, and *Streptococcus* strains were related to the production of the amyloid fibers.<sup>230</sup> The bacterial amyloids could pass through the gut barrier and access the circulation resulting in increasing the secretion of pro-inflammatory cytokines, such as IL17A and IL-22. These proteins and cytokines could pass through the blood-brain-barrier (BBB) to reach the brain and promote the activation of NF- $\kappa$ B, which provided neuroinflammation and the upregulation of miRNA-34a. miRNA-34a promotes the downregulation of the triggering receptor expressed in the microglial/myeloid cells-2 (TREM2) expression leading to the impairment of phagocytosis and the accumulation of amyloid proteins in the brain, which is an important risk factor to AD (Fig. 2).<sup>236,238–240</sup> Another microbial metabolite





that could contribute to neurological diseases is TMAO, which has been found elevated in the cerebrospinal fluid (CSF) of AD patients and associated with an important Alzheimer biomarker, p-tau protein,<sup>26</sup> so that the reduction of TMAO would work as a therapeutic tool to improve cognitive impairment.<sup>86</sup>

The ability of metformin to elevate anti-inflammatory SCFA-producing strains, such as Roseburia, which in turn were found in low concentrations in AD patients, was also demonstrated.<sup>99</sup> In addition, metformin promoted the reduction of the TMAO levels by increasing the strains negatively related to the production of TMAO (Akkermansia and Bifidobacterium) and reducing strains associated with TMAO production, such as Lachnospirillum and Ruminoclostridium.<sup>215</sup> Therefore, by reducing the plasma TMAO levels and enhancing the SCFAs, metformin would exert an anti-inflammatory action and contribute to preventing the progression of AD.

Amyloid fiber-producing strains are considered a concern for AD. In this context, metformin has an antimicrobial action against amyloid fibers producers, such as the Salmonella, Staphylococcus aureus, and Mycobacterium strains.<sup>241,242</sup> Beyond its action on microbiota, metformin through the AMPK activation is able to reduce the p62 accumulation in microglia (an indicator of the impairment of the autophagy process) leading to an increase in the phagocytosis of the A $\beta$  plaques and tau proteins by microglia, consequently reducing the brain deposits of these proteins that corroborated to an anti-AD effect of metformin.<sup>243</sup>

Furthermore, obesity is a risk factor for the impairment of the cognitive function and for diseases like AD.<sup>237,244</sup> In this context, a study by Ma *et al.* demonstrated that metformin was able to restore the cognitive impairment related to obesity through the recuperation of neurogenesis by the modulation of microbiota that acted on the attenuation of neuroinflammation.<sup>245</sup> Moreover, this study corroborated the observation that through its action in pathologies at risk for AD, such as diabetes and obesity,<sup>246</sup> where metformin promoted the increase of butyrate-producing strains that were associated with intestinal integrity.<sup>7</sup> It could prevent bacterial products, such as LPS from accessing the host's circulatory system and causing systemic inflammation, which could favor neuroinflammation and the development of AD.<sup>106,247</sup>

### Metformin and intestinal ischemia-reperfusion (I/R) injury

Intestinal ischemia reperfusion injury is a surgical complication characterized by the temporary interruption of blood circulation in the intestine, which is a serious complication that could lead to death. During the ischemia, the gastrointestinal barrier is impaired possibly from the release of microorganisms and of inflammatory factors. When the reperfusion occurs, these microorganisms can access the blood circulation and initiate an inflammatory cascade and sepsis.<sup>248,249</sup>

Understanding that the ischemia is an unpredictable complication, adopting strategies during the reperfusion that would reduce the inflammation and cellular damage would be important. In this context, the study of Jia *et al.* applied metformin before the reperfusion in mice, which resulted in the protection of the gastrointestinal barrier and prevented the ZO-1 and occludin tight junctions' damage. Beyond that, it was observed that metformin reduced (1) inflammatory factors, such as IL-6 and IL-1 $\beta$ , (2) the activation of NLRP3 inflammasome, (3) cleaved caspase-1, and (4) the levels of gasdermin D (GSDMD). These were important factors for pyroptosis, an inflammatory cell death predominant during the I/R. Thus, metformin proved to be a promising therapeutic tool for intestinal I/R injury; however, its clinical application would be

necessary for evaluating metformin's effects on intestinal I/R injury in humans.<sup>250</sup>

### Metformin and its role in healthy aging

Aging can be understood as a process that people go through as they get older, which is determined by genetics and influenced by the environment.<sup>251</sup> Associated with the improvement in the quality of life, people are living longer; as a consequence, there is an increasing number of people that are going through aging. The WHO's data has estimated that between 2015–2050, the proportion of people who will reach the age of 60 years or more, in relation to the world population, will increase from 12% to 22%.<sup>252</sup> In addition, it is estimated that between 2020–2050, the number of people aged 80 years or more will triple and comprise more than 426 million people worldwide.<sup>252</sup>

The molecular and cellular characteristics of aging involve (1) genomic instability, (2) telomeric wear, (3) epigenetic changes, (4) loss of proteases, (5) mitochondrial impairment, (6) cellular senescence, and (7) impairment of stem cells and intercellular communication.<sup>253</sup> Nevertheless, studies have pointed to the influence of intestinal microbiota on healthy aging.<sup>1,254</sup>

During aging, the gradual impairment of immunity is commonly associated with the loss of balance between pro- and anti-inflammatory actions.<sup>255</sup> Thus, aging is marked by a chronic inflammation called inflammaging, an important risk factor for the development of chronic diseases that increase the chance of mortality in these individuals like CVDs, dementia, and diabetes mellitus.<sup>256–259</sup> Thus, the reduction of inflammatory processes enables healthy aging.

By characterizing the microbiota of older individuals, it was possible to highlight the reduction of strains associated with butyrate production (Table 2).<sup>260–270</sup> The reduction of SCFAs was associated to the AMPK inhibition that promoted the increase of gut permeability and possibly the LPS release in circulation. The LPS was recognized by TLR4 in the macrophages surface resulting in macrophage polarization to the M1 phenotype and in an associated-inflammatory cascade that involved NF- $\kappa$ B activation and secretion of pro-inflammatory cytokines, consequently producing an inflammatory environment during aging and increasing the risk of the development of diseases that could compromise healthy aging.<sup>205,271</sup>

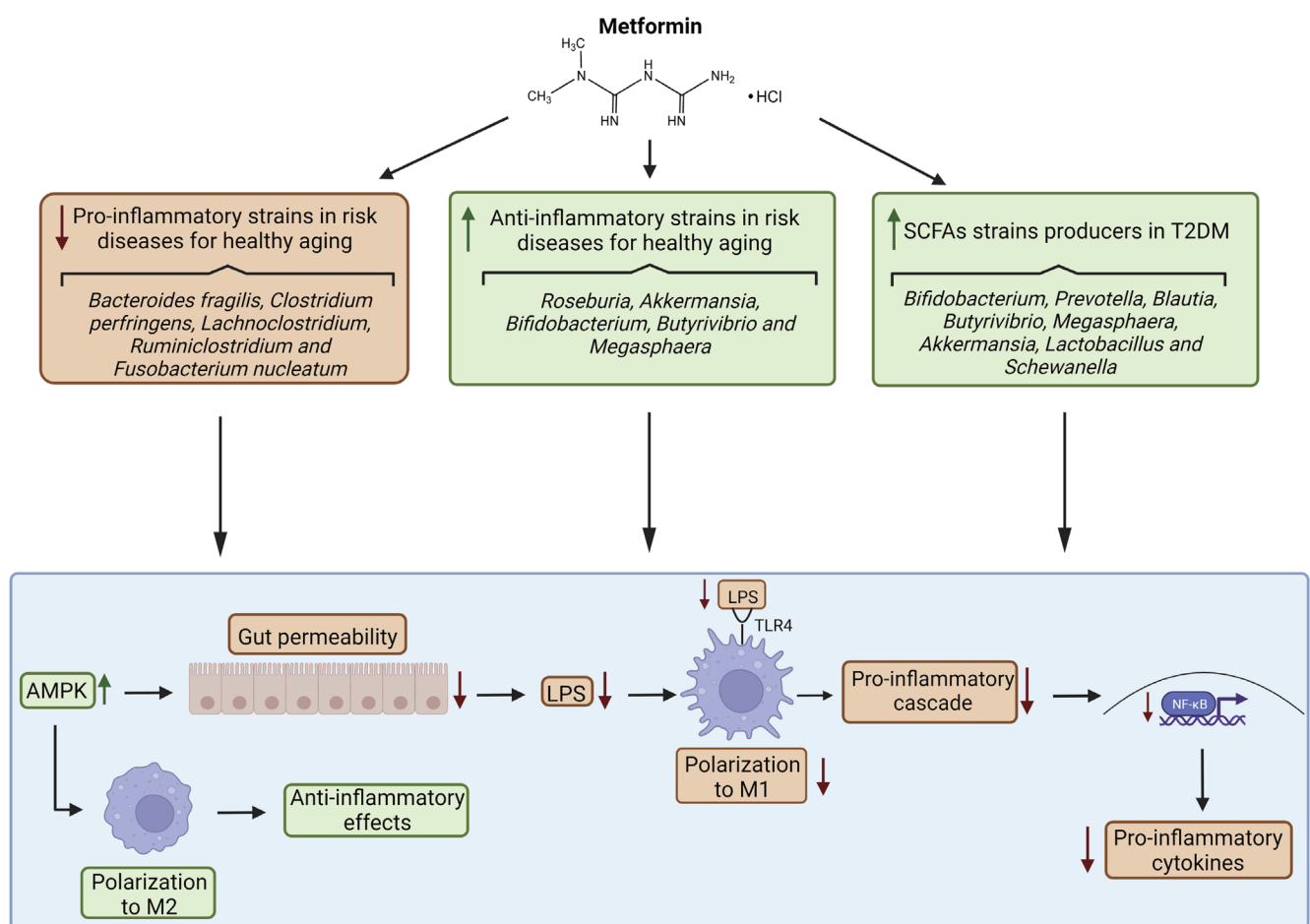
Metformin is associated with long and healthy aging,<sup>272–274</sup> due to its antidiabetic,<sup>263,264</sup> cardioprotective,<sup>265,275</sup> anti-tumor,<sup>266,267</sup> neuroprotective action,<sup>276</sup> and through its action on the microbiota.<sup>277,278</sup> Deficiency on SCFA-producing strains in elderly indicates a scenario of high intestinal permeability and inflammation.<sup>279,280</sup> Ahmadi *et al.* demonstrated these features among older mice that were reduced by metformin by increasing mucin production in the gastrointestinal tract.<sup>281</sup> Metformin increased the abundance of the strains associated with the production of the SCFAs in diabetic patients, such as Bifidobacterium, Prevotella, Blautia, Butyrivibrio, Megasphaera, Akkermansia, Lactobacillus, and Shewanella,<sup>56,167,207</sup> hence reducing the inflammatory environment and contributing to healthy aging (Fig. 3).

In addition, metformin was able to modulate the microbiota to exert its cardioprotective, anti-obesity, antidiabetic, antitumor, and neuroprotective effects, which in turn were considered risk factors for long-term healthy aging.<sup>282–287</sup> In this way, the microbial modulation exerted by metformin led to the effects listed above by reducing the pro-inflammatory and increasing the anti-inflammatory strains and consequently acting on aging (Fig. 3).<sup>56,138,167,215,287</sup>

**Table 2. Dysbiosis related to the aging process**

Authors	Results
Kim et al.; <sup>263</sup> Wu et al.; <sup>264</sup> Odamaki et al.; <sup>268</sup> Yu et al.; <sup>269</sup> Rizzatti et al. <sup>270</sup>	↑Proteobacteria (gram negative strain and LPS producer related to inflammation)
Biagi et al. <sup>265</sup>	↓Families Bacteroidaceae, Lachnospiraceae and Ruminococcaceae; ↑Genus Eggerthella, Akkermansia, Anaerotruncus, and Bilophila
Kushugulova et al. <sup>266</sup>	↓Butyricimonas virosa and Anaerostipes butyraticus (Butyrate producers strains related to anti-inflammatory properties)
Drago et al.; <sup>267</sup> Wu et al. <sup>264</sup>	↑Clostridia sensu stricto, Methanobrevibacter smithii and Bifidobacterium adolescentes (anti-inflammatory strain); ↓Faecalibacterium prausnitzii*, Dorea longicatena, Eubacterium rectale*, Bacteroides caccae and Fusobacterium mortiferum; * SCFAs strains producers related to anti-inflammatory properties and reduced risk for the development of inflammation-related diseases

LPS, lipopolysaccharide; SCFAs, short chain fatty acids.



**Fig. 3. Metformin and the anti-aging effects through microbiota modulation.** Metformin enhanced the SCFAs strains (Bifidobacterium, Prevotella, Blautia, Butyrivibrio, Megasphaera, Akkermansia, Lactobacillus, and Schewanella) in T2DM patients. Through its action against cancer, CVDs, obesity, AD, and diabetes (risk factors for healthy aging), metformin reduces pro-inflammatory strains (Bacteroides fragilis, Clostridium perfringens, Lachnoclostridium, Ruminiclostridium, and Fusobacterium nucleatum) and enhances anti-inflammatory strains (Roseburia, Akkermansia, Bifidobacterium, Butyrivibrio, and Megasphaera). Thus, through microbiota modulation, metformin promoted the activation of AMPK resulting in the reduction of gut permeability, reducing LPS in circulation and the activation of macrophage (M1 phenotype) through LPS recognition by TLR4 by avoiding (1) a pro-inflammatory cascade, (2) the activation of NF-κB, and (3) the production of pro-inflammatory cytokines. The AMPK activation was also related to macrophage polarization to the M2 phenotype, which had anti-inflammatory properties. Therefore, metformin reduced inflammation and the development of inflammation-related diseases contributing to its anti-aging effect. AD, Alzheimer disease; AMPK, AMP-activated protein kinase; CVDs, cardiovascular diseases; LPS, lipopolysaccharide; NF-κB, nuclear factor kappa B; SCFAs, short chain fatty acid; T2DM, type 2 diabetes mellitus; TLR4, toll-like receptor 4. Created with www.biorender.com.

### Metformin pharmacogenomics and the response to the drug

Metformin is a hydrophilic drug and a cation at physiological pH, so it is unable to cross the plasma membrane passively and requires transporters to enter for its excretion.<sup>288</sup> Several transporters are involved in the transporting of metformin that are tissue-specific, but those that play an important role in defining a response to metformin are (1) organic cation transporters 1–3 (OCTs), (2) the plasma membrane monoamine transporter (PMAT), (3) the protein of multidrug and toxin extrusion 1- 2 (MATE), (4) the serotonin transporter (SERT), and (5) the high-affinity choline transporter (CHT).<sup>31</sup> Despite the variety of transporters, the main ones are the OCTs, PMATs, and MATEs that play an important role in defining a response to metformin.<sup>31,289</sup>

In addition, entry by metformin into the gastrointestinal (GI) tract is crucial for its action. The transporting of metformin to the enterocytes can be mediated by different transporters, such as OCT1, OCT3, PMAT, SERT, and CHT; however, *in vitro* and *in vivo* studies have shown that the main mediators of transporting metformin to the enterocytes are OCT1, PMAT, and SERT.<sup>31,288,289</sup> Subsequently, metformin enters the bloodstream and OCT1 mediates the liver uptake, where it would play its main mechanisms of action. Moreover, the excretion of metformin involves two steps: (1) Its transport from the blood to the kidneys via OCT2 and (2) renal extrusion mediated by the MATE1 and 2 transporters, thus enabling its elimination in the urine.<sup>290</sup>

Despite its transporters, about 30% of administered metformin is not absorbed from the GI tract and is eliminated in the feces.<sup>291</sup> Therefore, its uptake by the enterocytes is considered the limiting step to determine its bioavailability, which could vary from 20–70% between individuals.<sup>292–294</sup> In this context, studies have indicated that gene polymorphisms of the metformin transporters could compromise their efficiency, thereby resulting in a lower uptake and response to metformin.<sup>268,290,295,296</sup>

#### OCT1 (SLC22A1) polymorphisms

The SLC22A1 gene responsible for encoding the OCT1 transporter is highly susceptible to polymorphisms that compromise its function.<sup>291</sup> Several studies have already identified different polymorphisms in this gene in different populations, which is an observation of concern given that OCT1 is considered the central transporter for metformin activity and impacts its antihyperglycemic action.<sup>269,270</sup>

The study by Seitz *et al.* covered 52 countries and identified 85 OCT1 variants in different populations.<sup>297</sup> The main polymorphisms observed were (1) rs628031 (A<G) that is considered a very common variant and associated with the reduced expression of OCT1 in enterocytes, and promotes a lower uptake of metformin and its accumulation,<sup>298</sup> (2) rs122083571, a polymorphism associated with lower OCT1 activity,<sup>299–301</sup> (3) rs72552763, associated with a reduced uptake of metformin (>60%),<sup>297,302</sup> and (4) rs34059508, associated with compromised OCT1 localization and its inactivation.<sup>302</sup>

In addition to the importance of OCT1 for the antihyperglycemic action of metformin, the study by Sam *et al.* demonstrated the role of OCT1 in the anti-obesity action of metformin. In this study, it was observed that variations in the SLC22A1 gene would be associated with a lower uptake of metformin by the adipose tissue and the consequent lower inhibition of lipid accumulation by metformin.<sup>270</sup> Thus, the polymorphisms observed in the SLC22A1 gene could explain why 35% of T2DM patients did not respond to metformin, and indicating the importance of OCT1 for the response to this drug.<sup>268</sup>

#### OCT2 (SLC22A2)

The SLC22A2 gene responsible for encoding the OCT2 transporter plays an important role in the renal excretion of metformin; thus, variations in this gene have been related to impaired renal excretion of this drug.<sup>294</sup> More than 500 variations of SLC22A2 have already been identified. Among the variations found, the following stand out: (1) p.165M>I, p.199T>I, p.201T>M, and p.400R>C, which are associated with lower OCT2 activity<sup>303,304</sup> and (2) p.270A>S (c.808G>T), which has been identified in different populations and is associated with a 40% reduction in metformin excretion in patients with the homozygous genotype.<sup>305</sup>

Impaired renal excretion of metformin is also associated with variations of metformin concentration in plasma between patients and adverse effects, such as elevated hypoglycemia.<sup>294</sup>

#### MATE1 (SLC47A1)/MATE2 (SLC47A2)

The multidrug and toxin extrusion protein 1 (MATE1) is encoded by the SLC47A1 gene. It is highly expressed in the liver and kidney, and has metformin, among other drugs as a substrate. MATE 1 may excrete metformin from the hepatocytes,<sup>306</sup> and along with MATE 2 in the kidneys, is responsible for the extrusion of metformin into urine. *In vitro* studies have shown that some genomic variants of this gene induced a complete loss of function for the transportation of metformin among other drugs, while others significantly altered the transport function in a substrate dependent way.<sup>307</sup> These alterations in expression of MATE1 could lead to increased systemic metformin and lactic acidosis induced by metformin.<sup>308,309</sup>

Furthermore, the SLC47A2 gene responsible for encoding the MATE2 transporter is highly expressed in renal cells and acts in the renal excretion of metformin together with MATE1 and OCT2. MATE2 pharmacogenomics demonstrate that responses to metformin transport capabilities resulting from different genetic alterations, where some polymorphisms have been thought to impact negatively metformin extrusion,<sup>310</sup> while others impact positively, as the most common promoter variant of MATE2, g.-130G>A, when in homozygosity leads to a significant enhancement of metformin extrusion and renal clearance.<sup>311</sup>

#### LKB1 (STK11)

In addition to the transporters for the response to metformin, another important factor would be serine-threonine kinase 11 (LKB1) (STK11 gene), which participates in the activation of AMPK, through which metformin exerts most of its mechanisms of action.<sup>312</sup> In the study by Li *et al.*, it was observed that the rs2075604 variation was associated with an improvement in the response to metformin being able to promote the reduction of HbA1c and FBG (fasting blood glucose) more efficiently, which was important for the action of metformin in T2DM.

#### Future Directions

In the past years, many studies about the relationship of metformin, microbiota, and diseases have been performed and bringing important discoveries and updates. Through these studies, it was possible to observe the important role of the microbiota in the development, progression, and treatment response of many diseases, in addition to the importance of metformin as a microbiota regulator. Understanding the importance of a regulated microbiota for health, its use as a therapeutic tool through a fecal microbiota transplan-

tation (FMT) has been growing. Hu *et al.* demonstrated that the FMT in a myocarditis mouse model improved the symptoms of the disease.<sup>313</sup> Likewise, in the study of Wang *et al.*, it showed a reduction of hyperglycemia, glycated hemoglobin, and inflammation beyond the improvement of insulinemia and insulin sensitivity after a FMT of a healthy mouse to a T2DM mouse model.<sup>314</sup> Additionally, Broadfield *et al.* were the first to shown an FMT from a metformin-treated healthy mouse to a CRC mouse model that was responsible for reducing the tumor volume and cholesterol metabolism, in addition to the enhancement of the SCFAs producers,<sup>315</sup> a transplant methodology that would be beneficial for individuals with reduced response to metformin due to metformin's transporters polymorphisms. This would indicate that the FMT would be a promising therapeutic tool, and its use combined with metformin could promote better results.

### Conclusions

The importance of the microbiota for the health of the host and its involvement with several pathologies have been studied with different perspectives. It was found that most of them converged to the central role of dysbiosis in the development of several pathologies, and highlighted the importance of therapeutic agents capable of regulating the host microbiota by exploiting microbiota's potential role in prophylaxis and treatment. The role of metformin in the regulation of the microbiota also has a strong impact on the prevention and progression of pathologies, such as obesity, T2DM, colorectal cancer, HIV, CVDs and in the healthy aging process. Pharmacogenomic data on metformin absorption and excretion added an important tool to predict patients' responses to the presence of metformin, and how this could impact microbiota. The role of probiotics and prebiotics as adjuvants to the antidiabetic action of metformin through the modulation of the intestinal microbiota has been widely studied, while its impact on HIV infection<sup>316</sup> and neurodegenerative diseases<sup>317</sup> is still under debate, thus more studies would be necessary. The understanding of the interplay of so many factors on patients' outcome for several diseases is highly complex, and more correlations should be explored.

### Acknowledgments

None.

### Funding

The present study was supported by the Research Support Foundation of Rio de Janeiro (FAPERJ; grant no. E-26/201.113/2019).

### Conflict of interest

Dr. Janaina Fernandes has been an editorial board member of *Journal of Exploratory Research in Pharmacology* since November 2021. The authors have no other conflicts of interest to declare.

### Author contributions

Contributed to the study concept and design (GSP and JF), acquisition of the data (GSP and JF), assay performance and data analysis (GSP and JF), drafting of the manuscript (GSP), critical revision of the manuscript (JF), and supervision (JF).

### References

- [1] Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. *Cell* 2012;148(6):1258–1270. doi:10.1016/j.cell.2012.01.035, PMID:22424233.
- [2] Lynch SV, Pedersen O. The Human Intestinal Microbiome in Health and Disease. *N Engl J Med* 2016;375(24):2369–2379. doi:10.1056/NEJMr1600266, PMID:27974040.
- [3] Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, *et al.* MetaHIT Consortium. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 2010;464(7285):59–65. doi:10.1038/nature08821, PMID:20203603.
- [4] Bäckhed F, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI. Host-bacterial mutualism in the human intestine. *Science* 2005;307(5717):1915–1920. doi:10.1126/science.1104816, PMID:15790844.
- [5] Pascale A, Marchesi N, Govoni S, Coppola A, Gazzaruso C. The role of gut microbiota in obesity, diabetes mellitus, and effect of metformin: new insights into old diseases. *Curr Opin Pharmacol* 2019;49:1–5. doi:10.1016/j.coph.2019.03.011, PMID:31015106.
- [6] Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. *Nature* 2012;486(7402):207–214. doi:10.1038/nature11234, PMID:22699609.
- [7] Iulia-Suceveanu A, Ioan Micu S, Voinea C, Elena Manea M, Catrinou D, Mazilu L, *et al.* Metformin and its benefits in improving gut microbiota disturbances in diabetes patients. In: Stoian AMP, Rizzo M (eds). *Metformin* [Internet]. London: IntechOpen; 2019. doi:10.5772/intechopen.88749.
- [8] Pascale A, Marchesi N, Marelli C, Coppola A, Luzi L, Govoni S, *et al.* Microbiota and metabolic diseases. *Endocrine* 2018;61(3):357–371. doi:10.1007/s12020-018-1605-5, PMID:29721802.
- [9] DeGruttola AK, Low D, Mizoguchi A, Mizoguchi E. Current Understanding of Dysbiosis in Disease in Human and Animal Models. *Inflamm Bowel Dis* 2016;22(5):1137–1150. doi:10.1097/MIB.0000000000000750, PMID:27070911.
- [10] Bailey CJ. Metformin: historical overview. *Diabetologia* 2017; 60(9):1566–1576. doi:10.1007/s00125-017-4318-z, PMID:28776081.
- [11] Blonde L, Dipp S, Cadena D. Combination Glucose-Lowering Therapy Plans in T2DM: Case-Based Considerations. *Adv Ther* 2018;35(7):939–965. doi:10.1007/s12325-018-0694-0, PMID:29777519.
- [12] Lv Z, Guo Y. Metformin and Its Benefits for Various Diseases. *Front Endocrinol (Lausanne)* 2020;11:191. doi:10.3389/fendo.2020.00191, PMID:32425881.
- [13] Choi YK, Park KG. Metabolic roles of AMPK and metformin in cancer cells. *Mol Cells* 2013;36(4):279–287. doi:10.1007/s10059-013-0169-8, PMID:23794020.
- [14] Demaré S, Kothari A, Calcutt NA, Fernyhough P. Metformin as a potential therapeutic for neurological disease: mobilizing AMPK to repair the nervous system. *Expert Rev Neurother* 2021;21(1):45–63. doi:10.1080/14737175.2021.1847645, PMID:33161784.
- [15] Luo T, Nocon A, Fry J, Sherban A, Rui X, Jiang B, *et al.* AMPK Activation by Metformin Suppresses Abnormal Extracellular Matrix Remodeling in Adipose Tissue and Ameliorates Insulin Resistance in Obesity. *Diabetes* 2016;65(8):2295–2310. doi:10.2337/db15-1122, PMID:27207538.
- [16] Viollet B, Guigas B, Sanz Garcia N, Leclerc J, Foretz M, Andreelli F. Cellular and molecular mechanisms of metformin: an overview. *Clin Sci (Lond)* 2012;122(6):253–270. doi:10.1042/CS20110386, PMID:22117616.
- [17] Hardie DG, Ross FA, Hawley SA. AMP-activated protein kinase: a target for drugs both ancient and modern. *Chem Biol* 2012;19(10):1222–1236. doi:10.1016/j.chembiol.2012.08.019, PMID:23102217.
- [18] Zhang Q, Hu N. Effects of Metformin on the Gut Microbiota in Obesity and Type 2 Diabetes Mellitus. *Diabetes Metab Syndr Obes* 2020;13:5003–5014. doi:10.2147/DMSO.S286430, PMID:33364804.
- [19] Olokoba AB, Obateru OA, Olokoba LB. Type 2 diabetes mellitus: a review of current trends. *Oman Med J* 2012;27(4):269–273. doi:10.5001/omj.2012.68, PMID:23071876.
- [20] Khan MAB, Hashim MJ, King JK, Govender RD, Mustafa H, Al Kaabi J. Epidemiology of Type 2 Diabetes - Global Burden of Disease and Forecasted Trends. *J Epidemiol Glob Health* 2020;10(1):107–111. doi:10.2991/jegh.k.191028.001, PMID:32175717.
- [21] Pearson ER. Type 2 diabetes: a multifaceted disease. *Diabetologia* 2019;

- 62(7):1107–1112. doi:10.1007/s00125-019-4909-y, PMID:31161345.
- [22] Wang Y, Luo X, Mao X, Tao Y, Ran X, Zhao H, *et al*. Gut microbiome analysis of type 2 diabetic patients from the Chinese minority ethnic groups the Uygurs and Kazaks. *PLoS One* 2017;12(3):e0172774. doi:10.1371/journal.pone.0172774, PMID:28328990.
- [23] Gurung M, Li Z, You H, Rodrigues R, Jump DB, Morgun A, *et al*. Role of gut microbiota in type 2 diabetes pathophysiology. *EBioMedicine* 2020;51:102590. doi:10.1016/j.ebiom.2019.11.051, PMID:31901868.
- [24] Harsch IA, Konturek PC. The Role of Gut Microbiota in Obesity and Type 2 and Type 1 Diabetes Mellitus: New Insights into “Old” Diseases. *Med Sci (Basel)* 2018;6(2):E32. doi:10.3390/medsci6020032, PMID:29673211.
- [25] Aguirre M, Eck A, Koenen ME, Savelkoul PH, Budding AE, Venema K. Diet drives quick changes in the metabolic activity and composition of human gut microbiota in a validated in vitro gut model. *Res Microbiol* 2016;167(2):114–125. doi:10.1016/j.resmic.2015.09.006, PMID:26499094.
- [26] Larsen N, Vogensen FK, van den Berg FW, Nielsen DS, Andreasen AS, Pedersen BK, *et al*. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. *PLoS One* 2010;5(2):e9085. doi:10.1371/journal.pone.0009085, PMID:20140211.
- [27] Bailey CJ, Wilcock C, Scarpello JH. Metformin and the intestine. *Diabetologia* 2008;51(8):1552–1553. doi:10.1007/s00125-008-1053-5, PMID:18528677.
- [28] Karlsson F, Tremaroli V, Nielsen J, Bäckhed F. Assessing the human gut microbiota in metabolic diseases. *Diabetes* 2013;62(10):3341–3349. doi:10.2337/db13-0844, PMID:24065795.
- [29] Jones GR, Molloy MP. Metformin, Microbiome and Protection Against Colorectal Cancer. *Dig Dis Sci* 2021;66(5):1409–1414. doi:10.1007/s10620-020-06390-4, PMID:32533543.
- [30] Shin NR, Lee JC, Lee HY, Kim MS, Whon TW, Lee MS, *et al*. An increase in the Akkermansia spp. population induced by metformin treatment improves glucose homeostasis in diet-induced obese mice. *Gut* 2014;63(5):727–735. doi:10.1136/gutjnl-2012-303839, PMID:23804561.
- [31] McCreight LJ, Bailey CJ, Pearson ER. Metformin and the gastrointestinal tract. *Diabetologia* 2016;59(3):426–435. doi:10.1007/s00125-015-3844-9, PMID:26780750.
- [32] Ma W, Chen J, Meng Y, Yang J, Cui Q, Zhou Y. Metformin Alters Gut Microbiota of Healthy Mice: Implication for Its Potential Role in Gut Microbiota Homeostasis. *Front Microbiol* 2018;9:1336. doi:10.3389/fmicb.2018.01336, PMID:29988362.
- [33] Koh SJ, Kim JM, Kim IK, Ko SH, Kim JS. Anti-inflammatory mechanism of metformin and its effects in intestinal inflammation and colitis-associated colon cancer. *J Gastroenterol Hepatol* 2014;29(3):502–510. doi:10.1111/jgh.12435, PMID:24716225.
- [34] Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol* 2015;11(10):577–591. doi:10.1038/nrendo.2015.128, PMID:26260141.
- [35] De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchamp A, *et al*. Microbiota-generated metabolites promote metabolic benefits via gut-brain neural circuits. *Cell* 2014;156(1-2):84–96. doi:10.1016/j.cell.2013.12.016, PMID:24412651.
- [36] Sun L, Xie C, Wang G, Wu Y, Wu Q, Wang X, *et al*. Gut microbiota and intestinal FXR mediate the clinical benefits of metformin. *Nat Med* 2018;24(12):1919–1929. doi:10.1038/s41591-018-0222-4, PMID:30397356.
- [37] Newgard CB, An J, Bain JR, Muehlbauer MJ, Stevens RD, Lien LF, *et al*. A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metab* 2009;9(4):311–326. doi:10.1016/j.cmet.2009.02.002, PMID:19356713.
- [38] Sonnet DS, O’Leary MN, Gutierrez MA, Nguyen S, Mateen S, Hsu Y, *et al*. Metformin inhibits Branched Chain Amino Acid (BCAA) derived ketoacidosis and promotes metabolic homeostasis in MSUD. *Sci Rep* 2016;6:28775. doi:10.1038/srep28775, PMID:27373929.
- [39] Amin S, Lux A, O’Callaghan F. The journey of metformin from glycaemic control to mTOR inhibition and the suppression of tumour growth. *Br J Clin Pharmacol* 2019;85(1):37–46. doi:10.1111/bcp.13780, PMID:30290005.
- [40] Nadkarni P, Chepurny OG, Holz GG. Regulation of glucose homeostasis by GLP-1. *Prog Mol Biol Transl Sci* 2014;121:23–65. doi:10.1016/B978-0-12-800101-1.00002-8, PMID:24373234.
- [41] Zhao L, Chen Y, Xia F, Abudukerimu B, Zhang W, Guo Y, *et al*. A Glucagon-Like Peptide-1 Receptor Agonist Lowers Weight by Modulating the Structure of Gut Microbiota. *Front Endocrinol (Lausanne)* 2018;9:233. doi:10.3389/fendo.2018.00233, PMID:29867765.
- [42] Tsai CY, Lu HC, Chou YH, Liu PY, Chen HY, Huang MC, *et al*. Gut Microbial Signatures of Glycemic Responses of GLP-1 Receptor Agonists in Type 2 Diabetic Patients: A Pilot Study. *Front Endocrinol (Lausanne)* 2021;12:814770. doi:10.3389/fendo.2021.814770, PMID:35095773.
- [43] Koh A, Molinaro A, Ståhlman M, Khan MT, Schmidt C, Mannerås-Holm L, *et al*. Microbially Produced Imdazole Propionate Impairs Insulin Signaling through mTORC1. *Cell* 2018;175(4):947–961.e17. doi:10.1016/j.cell.2018.09.055, PMID:30401435.
- [44] Pryor R, Norvaisas P, Marinos G, Best L, Thingholm LB, Quintaneiro LM, *et al*. Host-Microbe-Drug-Nutrient Screen Identifies Bacterial Effectors of Metformin Therapy. *Cell* 2019;178(6):1299–1312.e29. doi:10.1016/j.cell.2019.08.003, PMID:31474368.
- [45] Bailey CJ, Wilcock C, Day C. Effect of metformin on glucose metabolism in the splanchnic bed. *Br J Pharmacol* 1992;105(4):1009–1013. doi:10.1111/j.1476-5381.1992.tb09093.x, PMID:1504710.
- [46] Koffert JP, Mikkola K, Virtanen KA, Andersson AD, Faxius L, Hällsten K, *et al*. Metformin treatment significantly enhances intestinal glucose uptake in patients with type 2 diabetes: Results from a randomized clinical trial. *Diabetes Res Clin Pract* 2017;131:208–216. doi:10.1016/j.diabres.2017.07.015, PMID:28778047.
- [47] Gorboulev V, Schürmann A, Vallon V, Kipp H, Jaschke A, Klessen D, *et al*. Na(+)-D-glucose cotransporter SGLT1 is pivotal for intestinal glucose absorption and glucose-dependent incretin secretion. *Diabetes* 2012;61(1):187–196. doi:10.2337/db11-1029, PMID:22124465.
- [48] Bauer PV, Duca FA, Waise TMZ, Rasmussen BA, Abraham MA, Dranse HJ, *et al*. Metformin Alters Upper Small Intestinal Microbiota that Impact a Glucose-SGLT1-Sensing Glucoregulatory Pathway. *Cell Metab* 2018;27(1):101–117.e5. doi:10.1016/j.cmet.2017.09.019, PMID:29056513.
- [49] Johns EC, Denison FC, Norman JE, Reynolds RM. Gestational Diabetes Mellitus: Mechanisms, Treatment, and Complications. *Trends Endocrinol Metab* 2018;29(11):743–754. doi:10.1016/j.tem.2018.09.004, PMID:30297319.
- [50] Mendez-Figueroa H, Schuster M, Maggio L, Pedroza C, Chauhan SP, Paglia MJ. Gestational Diabetes Mellitus and Frequency of Blood Glucose Monitoring: A Randomized Controlled Trial. *Obstet Gynecol* 2017;130(1):163–170. doi:10.1097/AOG.0000000000002101, PMID:28594772.
- [51] American Diabetes Association. 14. Management of Diabetes in Pregnancy: Standards of Medical Care in Diabetes-2020. *Diabetes Care* 2020(Suppl 1):S183–S192. doi:10.2337/dc20-S014, PMID:31862757.
- [52] Molina-Vega M, Picón-César MJ, Gutiérrez-Repiso C, Fernández-Valero A, Lima-Rubio F, González-Romero S, *et al*. Metformin action over gut microbiota is related to weight and glycemic control in gestational diabetes mellitus: A randomized trial. *Biomed Pharmacother* 2022;145:112465. doi:10.1016/j.biopha.2021.112465, PMID:34844107.
- [53] Shin NR, Gu N, Choi HS, Kim H. Combined effects of *Scutellaria baicalensis* with metformin on glucose tolerance of patients with type 2 diabetes via gut microbiota modulation. *Am J Physiol Endocrinol Metab* 2020;318(1):E52–E61. doi:10.1152/ajpendo.00221.2019, PMID:31770016.
- [54] Burton JH, Johnson M, Johnson J, Hsia DS, Greenway FL, Heiman ML. Addition of a Gastrointestinal Microbiome Modulator to Metformin Improves Metformin Tolerance and Fasting Glucose Levels. *J Diabetes Sci Technol* 2015;9(4):808–814. doi:10.1177/1932296815577425, PMID:25802471.
- [55] Palacios T, Vitetta L, Coulson S, Madigan CD, Lam YY, Manuel R, *et al*. Targeting the Intestinal Microbiota to Prevent Type 2 Diabetes and Enhance the Effect of Metformin on Glycaemia: A Randomised Controlled Pilot Study. *Nutrients* 2020;12(7):E2041. doi:10.3390/nu12072041, PMID:32660025.
- [56] de la Cuesta-Zuluaga J, Mueller NT, Corrales-Agudelo V, Velásquez-Mejía EP, Carmona JA, Abad JM, *et al*. Metformin Is Associated With Higher Relative Abundance of Mucin-Degrading *Akkermansia mucin-*

- iphila and Several Short-Chain Fatty Acid-Producing Microbiota in the Gut. *Diabetes Care* 2017;40(1):54–62. doi:10.2337/dc16-1324, PMID: 27999002.
- [57] Yan F, Li N, Yue Y, Wang C, Zhao L, Evivie SE, *et al*. Screening for Potential Novel Probiotics With Dipeptidyl Peptidase IV-Inhibiting Activity for Type 2 Diabetes Attenuation in vitro and in vivo. *Front Microbiol* 2019;10:2855. doi:10.3389/fmicb.2019.02855, PMID:31998245.
- [58] Haase CL, Eriksen KT, Lopes S, Satyrganova A, Schnecke V, McEwan P. Body mass index and risk of obesity-related conditions in a cohort of 2.9 million people: Evidence from a UK primary care database. *Obes Sci Pract* 2021;7(2):137–147. doi:10.1002/osp4.474, PMID: 33841883.
- [59] Passos CMD, Maia EG, Levy RB, Martins APB, Claro RM. Association between the price of ultra-processed foods and obesity in Brazil. *Nutr Metab Cardiovasc Dis* 2020;30(4):589–598. doi:10.1016/j.numecd.2019.12.011, PMID:32139251.
- [60] Chooi YC, Ding C, Magkos F. The epidemiology of obesity. *Metabolism* 2019;92:6–10. doi:10.1016/j.metabol.2018.09.005, PMID:30253139.
- [61] Hill JO. Understanding and addressing the epidemic of obesity: an energy balance perspective. *Endocr Rev* 2006;27(7):750–761. doi: 10.1210/er.2006-0032, PMID:17122359.
- [62] Clavel T, Desmarchelier C, Haller D, Gérard P, Rohn S, Lepage P, *et al*. Intestinal microbiota in metabolic diseases: from bacterial community structure and functions to species of pathophysiological relevance. *Gut Microbes* 2014;5(4):544–551. doi:10.4161/gmic.29331, PMID:25003516.
- [63] Rosenbaum M, Knight R, Leibel RL. The gut microbiota in human energy homeostasis and obesity. *Trends Endocrinol Metab* 2015;26(9):493–501. doi:10.1016/j.tem.2015.07.002, PMID:26257300.
- [64] Ley RE, Bäckhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut microbial ecology. *Proc Natl Acad Sci U S A* 2005;102(31):11070–11075. doi:10.1073/pnas.0504978102, PMID:16 033867.
- [65] Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature* 2006;444(7122):1022–1023. doi:10.1038/4441022a, PMID:17183309.
- [66] Armougom F, Henry M, Vialettes B, Raccach D, Raoult D. Monitoring bacterial community of human gut microbiota reveals an increase in *Lactobacillus* in obese patients and *Methanogens* in anorexic patients. *PLoS One* 2009;4(9):e7125. doi:10.1371/journal.pone.0007125, PMID:1977 4074.
- [67] Furet JP, Kong LC, Tap J, Poitou C, Basdevant A, Bouillot JL, *et al*. Differential adaptation of human gut microbiota to bariatric surgery-induced weight loss: links with metabolic and low-grade inflammation markers. *Diabetes* 2010;59(12):3049–3057. doi:10.2337/db10-0253, PMID:20876719.
- [68] Santacruz A, Collado MC, García-Valdés L, Segura MT, Martín-Lagos JA, Anjos T, *et al*. Gut microbiota composition is associated with body weight, weight gain and biochemical parameters in pregnant women. *Br J Nutr* 2010;104(1):83–92. doi:10.1017/S0007114510000176, PMID: 20205964.
- [69] Magne F, Gotteland M, Gauthier L, Zazueta A, Pesoa S, Navarrete P, *et al*. The Firmicutes/Bacteroidetes Ratio: A Relevant Marker of Gut Dysbiosis in Obese Patients? *Nutrients* 2020;12(5):E1474. doi:10.3390/nu12051474, PMID:32438689.
- [70] Schwirtz A, Taras D, Schäfer K, Beijer S, Bos NA, Donus C, *et al*. Microbiota and SCFA in lean and overweight healthy subjects. *Obesity (Silver Spring)* 2010;18(1):190–195. doi:10.1038/oby.2009.167, PMID:19498350.
- [71] Collado MC, Isolauri E, Laitinen K, Salminen S. Distinct composition of gut microbiota during pregnancy in overweight and normal-weight women. *Am J Clin Nutr* 2008;88(4):894–899. doi:10.1093/ajcn/88.4.894, PMID:18842773.
- [72] Duncan SH, Lobley GE, Holtrop G, Ince J, Johnstone AM, Louis P, *et al*. Human colonic microbiota associated with diet, obesity and weight loss. *Int J Obes (Lond)* 2008;32(11):1720–1724. doi:10.1038/ijo.2008.155, PMID:18779823.
- [73] Jumpertz R, Le DS, Turnbaugh PJ, Trinidad C, Bogardus C, Gordon JI, *et al*. Energy-balance studies reveal associations between gut microbes, caloric load, and nutrient absorption in humans. *Am J Clin Nutr* 2011;94(1):58–65. doi:10.3945/ajcn.110.010132, PMID:21543530.
- [74] Kalliomäki M, Collado MC, Salminen S, Isolauri E. Early differences in fecal microbiota composition in children may predict overweight. *Am J Clin Nutr* 2008;87(3):534–538. doi:10.1093/ajcn/87.3.534, PMID: 18326589.
- [75] Yin YN, Yu QF, Fu N, Liu XW, Lu FG. Effects of four Bifidobacteria on obesity in high-fat diet induced rats. *World J Gastroenterol* 2010;16(27):3394–3401. doi:10.3748/wjg.v16.i27.3394, PMID:20632441.
- [76] Gao R, Zhu C, Li H, Yin M, Pan C, Huang L, *et al*. Dysbiosis Signatures of Gut Microbiota Along the Sequence from Healthy, Young Patients to Those with Overweight and Obesity. *Obesity (Silver Spring)* 2018;26(2):351–361. doi:10.1002/oby.22088, PMID:29280312.
- [77] Koleva PT, Bridgman SL, Kozyrskij AL. The infant gut microbiome: evidence for obesity risk and dietary intervention. *Nutrients* 2015;7(4):2237–2260. doi:10.3390/nu7042237, PMID:25835047.
- [78] Dao MC, Everard A, Aron-Wisniewsky J, Sokolovska N, Prifti E, Verger EO, *et al*. MICRO-Obes Consortium. Akkermansia muciniphila and improved metabolic health during a dietary intervention in obesity: relationship with gut microbiome richness and ecology. *Gut* 2016;65(3):426–436. doi:10.1136/gutjnl-2014-308778, PMID:26100928.
- [79] Gérard P. Gut microbiota and obesity. *Cell Mol Life Sci* 2016;73(1):147–162. doi:10.1007/s00018-015-2061-5, PMID:26459447.
- [80] Khan MJ, Gerasimidis K, Edwards CA, Shaikh MG. Role of Gut Microbiota in the Aetiology of Obesity: Proposed Mechanisms and Review of the Literature. *J Obes* 2016;2016:7353642. doi:10.1155/2016/7353642, PMID:27703805.
- [81] Davis CD. The Gut Microbiome and Its Role in Obesity. *Nutr Today* 2016; 51(4):167–174. doi:10.1097/NT.000000000000167, PMID:27795585.
- [82] Zambelli K, Fitch MD, Fleming SE. Acetate and butyrate are the major substrates for de novo lipogenesis in rat colonic epithelial cells. *J Nutr* 2003; 133(11):3509–3515. doi:10.1093/jn/133.11.3509, PMID:14608066.
- [83] Roy CC, Kien CL, Bouthillier L, Levy E. Short-chain fatty acids: ready for prime time? *Nutr Clin Pract* 2006;21(4):351–366. doi:10.1177/011542 6506021004351, PMID:16870803.
- [84] Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, *et al*. The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *J Biol Chem* 2003;278(13):11312–11319. doi:10.1074/jbc.M211609200, PMID:12496283.
- [85] Cuevas-Ramos G, Petit CR, Marcq I, Boury M, Oswald E, Nougayrède JP. *Escherichia coli* induces DNA damage in vivo and triggers genomic instability in mammalian cells. *Proc Natl Acad Sci U S A* 2010;107 (25):11537–11542. doi:10.1073/pnas.1001261107, PMID:20534522.
- [86] Ellulu MS, Patimah I, Khaza'ai H, Rahmat A, Abed Y. Obesity and inflammation: the linking mechanism and the complications. *Arch Med Sci* 2017; 13(4):851–863. doi:10.5114/aoms.2016.58928, PMID:28721154.
- [87] Guo S, Al-Sadi R, Said HM, Ma TY. Lipopolysaccharide causes an increase in intestinal tight junction permeability in vitro and in vivo by inducing enterocyte membrane expression and localization of TLR-4 and CD14. *Am J Pathol* 2013;182(2):375–387. doi:10.1016/j.ajpath.2012.10.014, PMID:23201091.
- [88] Hiippala K, Jouhten H, Ronkainen A, Hartikainen A, Kainulainen V, Jalanka J, *et al*. The Potential of Gut Commensals in Reinforcing Intestinal Barrier Function and Alleviating Inflammation. *Nutrients* 2018;10(8):E988. doi:10.3390/nu10080988, PMID:30060606.
- [89] Kim KN, Yao Y, Ju SY. Short Chain Fatty Acids and Fecal Microbiota Abundance in Humans with Obesity: A Systematic Review and Meta-Analysis. *Nutrients* 2019;11(10):E2512. doi:10.3390/nu11102512, PMID:316 35264.
- [90] Chukir T, Mandel L, Tchang BG, Al-Mulla NA, Igel LI, Kumar RB, *et al*. Metformin-induced weight loss in patients with or without type 2 diabetes/prediabetes: A retrospective cohort study. *Obes Res Clin Pract* 2021;15(1):64–68. doi:10.1016/j.orcp.2020.12.005, PMID:33386253.
- [91] Kahn SE, Haffner SM, Heise MA, Herman WH, Holman RR, Jones NP, *et al*. ADOPT Study Group. Glycemic durability of rosiglitazone, metformin, or glyburide monotherapy. *N Engl J Med* 2006;355(23):2427–2443. doi:10.1056/NEJMoa066224, PMID:17145742.
- [92] Malin SK, Kashyap SR. Effects of metformin on weight loss: potential mechanisms. *Curr Opin Endocrinol Diabetes Obes* 2014;21(5):323–329. doi:10.1097/MED.000000000000095, PMID:25105996.
- [93] Seifarth C, Schehler B, Schneider HJ. Effectiveness of metformin on weight loss in non-diabetic individuals with obesity. *Exp Clin Endocrinol*

- Diabetes 2013;121(1):27–31. doi:10.1055/s-0032-1327734, PMID: 23147210.
- [94] Kim EK, Lee SH, Jhun JY, Byun JK, Jeong JH, Lee SY, *et al*. Metformin Prevents Fatty Liver and Improves Balance of White/Brown Adipose in an Obesity Mouse Model by Inducing FGF21. *Mediators Inflamm* 2016;2016:5813030. doi:10.1155/2016/5813030, PMID:27057099.
- [95] Breining P, Jensen JB, Sundelin EI, Gormsen LC, Jakobsen S, Busk M, *et al*. Metformin targets brown adipose tissue in vivo and reduces oxygen consumption in vitro. *Diabetes Obes Metab* 2018;20(9):2264–2273. doi:10.1111/dom.13362, PMID:29752759.
- [96] Yerevanian A, Soukas AA. Metformin: Mechanisms in Human Obesity and Weight Loss. *Curr Obes Rep* 2019;8(2):156–164. doi:10.1007/s13679-019-00335-3, PMID:30874963.
- [97] Mulherin AJ, Oh AH, Kim H, Grieco A, Lauffer LM, Brubaker PL. Mechanisms underlying metformin-induced secretion of glucagon-like peptide-1 from the intestinal L cell. *Endocrinology* 2011;152(12):4610–4619. doi:10.1210/en.2011-1485, PMID:21971158.
- [98] Hardie DG, Sakamoto K. AMPK: a key sensor of fuel and energy status in skeletal muscle. *Physiology (Bethesda)* 2006;21:48–60. doi:10.1152/physiol.00044.2005, PMID:16443822.
- [99] Steinberg GR, Carling D. AMP-activated protein kinase: the current landscape for drug development. *Nat Rev Drug Discov* 2019;18(7):527–551. doi:10.1038/s41573-019-0019-2, PMID:30867601.
- [100] Desjardins EM, Steinberg GR. Emerging Role of AMPK in Brown and Beige Adipose Tissue (BAT): Implications for Obesity, Insulin Resistance, and Type 2 Diabetes. *Curr Diab Rep* 2018;18(10):80. doi:10.1007/s11892-018-1049-6, PMID:30120579.
- [101] Ding L, Zhang F, Zhao MX, Ren XS, Chen Q, Li YH, *et al*. Reduced lipolysis response to adipose afferent reflex involved in impaired activation of adrenoceptor-cAMP-PKA-hormone sensitive lipase pathway in obesity. *Sci Rep* 2016;6:34374. doi:10.1038/srep34374, PMID:27694818.
- [102] van Dam AD, Kooijman S, Schilperoort M, Rensen PC, Boon MR. Regulation of brown fat by AMP-activated protein kinase. *Trends Mol Med* 2015;21(9):571–579. doi:10.1016/j.molmed.2015.07.003, PMID:26271143.
- [103] Zhang E, Jin L, Wang Y, Tu J, Zheng R, Ding L, *et al*. Intestinal AMPK modulation of microbiota mediates crosstalk with brown fat to control thermogenesis. *Nat Commun* 2022;13(1):1135. doi:10.1038/s41467-022-28743-5, PMID:35241650.
- [104] Lee H, Ko G. Effect of metformin on metabolic improvement and gut microbiota. *Appl Environ Microbiol* 2014;80(19):5935–5943. doi:10.1128/AEM.01357-14, PMID:25038099.
- [105] Cabreiro F, Au C, Leung KY, Vergara-Irigaray N, Cochemé HM, Noori T, *et al*. Metformin retards aging in *C. elegans* by altering microbial folate and methionine metabolism. *Cell* 2013;153(1):228–239. doi:10.1016/j.cell.2013.02.035, PMID:23540700.
- [106] Zhang X, Zhao Y, Xu J, Xue Z, Zhang M, Pang X, *et al*. Modulation of gut microbiota by berberine and metformin during the treatment of high-fat diet-induced obesity in rats. *Sci Rep* 2015;5:14405. doi:10.1038/srep14405, PMID:26396057.
- [107] Jing Y, Wu F, Li D, Yang L, Li Q, Li R. Metformin improves obesity-associated inflammation by altering macrophages polarization. *Mol Cell Endocrinol* 2018;461:256–264. doi:10.1016/j.mce.2017.09.025, PMID:28935544.
- [108] Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, *et al*. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* 2021;71(3):209–249. doi:10.3322/caac.21660, PMID:33538338.
- [109] Dekker E, Tanis PJ, Vleugels JLA, Kasi PM, Wallace MB. Colorectal cancer. *Lancet* 2019;394(10207):1467–1480. doi:10.1016/S0140-6736(19)32319-0, PMID:31631858.
- [110] Botteri E, Iodice S, Bagnardi V, Raimondi S, Lowenfels AB, Maisonneuve P. Smoking and colorectal cancer: a meta-analysis. *JAMA* 2008;300(23):2765–2778. doi:10.1001/jama.2008.839, PMID:19088354.
- [111] Cai S, Li Y, Ding Y, Chen K, Jin M. Alcohol drinking and the risk of colorectal cancer death: a meta-analysis. *Eur J Cancer Prev* 2014;23(6):532–539. doi:10.1097/CEJ.0000000000000076, PMID:25170915.
- [112] Kyrgiou M, Kalliala I, Markozannes G, Gunter MJ, Paraskevidis E, Gaba H, *et al*. Adiposity and cancer at major anatomical sites: umbrella review of the literature. *BMJ* 2017;356:j477. doi:10.1136/bmj.j477, PMID:28246088.
- [113] Chan DS, Lau R, Aune D, Vieira R, Greenwood DC, Kampman E, *et al*. Red and processed meat and colorectal cancer incidence: meta-analysis of prospective studies. *PLoS One* 2011;6(6):e20456. doi:10.1371/journal.pone.0020456, PMID:21674008.
- [114] Kostic AD, Chun E, Robertson L, Glickman JN, Gallini CA, Michaud M, *et al*. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. *Cell Host Microbe* 2013;14(2):207–215. doi:10.1016/j.chom.2013.07.007, PMID:23954159.
- [115] Nakatsu G, Li X, Zhou H, Sheng J, Wong SH, Wu WK, *et al*. Gut mucosal microbiome across stages of colorectal carcinogenesis. *Nat Commun* 2015;6:8727. doi:10.1038/ncomms9727, PMID:26515465.
- [116] Winawer SJ, Zauber AG, Ho MN, O'Brien MJ, Gottlieb LS, Sternberg SS, *et al*. Prevention of colorectal cancer by colonoscopic polypectomy. The National Polyp Study Workgroup. *N Engl J Med* 1993;329(27):1977–1981. doi:10.1056/NEJM199312303292701, PMID:8247072.
- [117] Imperiale TF, Glowinski EA, Lin-Cooper C, Larkin GN, Rogge JD, Ransohoff DF. Five-year risk of colorectal neoplasia after negative screening colonoscopy. *N Engl J Med* 2008;359(12):1218–1224. doi:10.1056/NEJMoa0803597, PMID:18799558.
- [118] Zell JA, Pelot D, Chen WP, McLaren CE, Gerner EW, Meyskens FL. Risk of cardiovascular events in a randomized placebo-controlled, double-blind trial of difluoromethylornithine plus sulindac for the prevention of sporadic colorectal adenomas. *Cancer Prev Res (Phila)* 2009;2(3):209–212. doi:10.1158/1940-6207.CAPR-08-0203, PMID:19258540.
- [119] Higurashi T, Nakajima A. Metformin and Colorectal Cancer. *Front Endocrinol (Lausanne)* 2018;9:622. doi:10.3389/fendo.2018.00622, PMID:30405532.
- [120] Shaw RJ, Lamia KA, Vasquez D, Koo SH, Bardeesy N, Depinho RA, *et al*. The kinase LKB1 mediates glucose homeostasis in liver and therapeutic effects of metformin. *Science* 2005;310(5754):1642–1646. doi:10.1126/science.1120781, PMID:16308421.
- [121] Tomimoto A, Endo H, Sugiyama M, Fujisawa T, Hosono K, Takahashi H, *et al*. Metformin suppresses intestinal polyp growth in *ApcMin/+* mice. *Cancer Sci* 2008;99(11):2136–2141. doi:10.1111/j.1349-7006.2008.00933.x, PMID:18803638.
- [122] Hosono K, Endo H, Takahashi H, Sugiyama M, Uchiyama T, Suzuki K, *et al*. Metformin suppresses azoxymethane-induced colorectal aberrant crypt foci by activating AMP-activated protein kinase. *Mol Carcinog* 2010;49(7):662–671. doi:10.1002/mc.20637, PMID:20564343.
- [123] Anisimov VN. Metformin for Prevention and Treatment of Colon Cancer: A Reappraisal of Experimental and Clinical Data. *Curr Drug Targets* 2016;17(4):439–446. doi:10.2174/1389450116666150309113305, PMID:25738299.
- [124] Chen W, Liu F, Ling Z, Tong X, Xiang C. Human intestinal lumen and mucosa-associated microbiota in patients with colorectal cancer. *PLoS One* 2012;7(6):e39743. doi:10.1371/journal.pone.0039743, PMID:22761885.
- [125] Cheng Y, Ling Z, Li L. The Intestinal Microbiota and Colorectal Cancer. *Front Immunol* 2020;11:615056. doi:10.3389/fimmu.2020.615056, PMID:33329610.
- [126] Coker OO, Nakatsu G, Dai RZ, Wu WKK, Wong SH, Ng SC, *et al*. Enteric fungal microbiota dysbiosis and ecological alterations in colorectal cancer. *Gut* 2019;68(4):654–662. doi:10.1136/gutjnl-2018-317178, PMID:30472682.
- [127] Bashir A, Miskeen AY, Bhat A, Fazili KM, Ganai BA. *Fusobacterium nucleatum*: an emerging bug in colorectal tumorigenesis. *Eur J Cancer Prev* 2015;24(5):373–385. doi:10.1097/CEJ.0000000000000116, PMID:25569450.
- [128] Liu X, Cheng Y, Shao L, Ling Z. Alterations of the Predominant Fecal Microbiota and Disruption of the Gut Mucosal Barrier in Patients with Early-Stage Colorectal Cancer. *Biomed Res Int* 2020;2020:2948282. doi:10.1155/2020/2948282, PMID:32280686.
- [129] Yang Y, Weng W, Peng J, Hong L, Yang L, Toyiyama Y, *et al*. *Fusobacterium nucleatum* Increases Proliferation of Colorectal Cancer Cells and Tumor Development in Mice by Activating Toll-Like Receptor 4 Signaling to Nuclear Factor- $\kappa$ B, and Up-regulating Expression of MicroRNA-21. *Gastroenterology* 2017;152(4):851–866.e24. doi:10.1053/j.gastro.2016.11.018, PMID:27876571.
- [130] Guo S, Li L, Xu B, Li M, Zeng Q, Xiao H, *et al*. A Simple and Novel Fecal Biomarker for Colorectal Cancer: Ratio of *Fusobacterium Nucleatum*

- to Probiotics Populations, Based on Their Antagonistic Effect. *Clin Chem* 2018;64(9):1327–1337. doi:10.1373/clinchem.2018.289728, PMID:29914865.
- [131] Mima K, Nishihara R, Qian ZR, Cao Y, Sukawa Y, Nowak JA, *et al*. *Fusobacterium nucleatum* in colorectal carcinoma tissue and patient prognosis. *Gut* 2016;65(12):1973–1980. doi:10.1136/gutjnl-2015-310101, PMID:26311717.
- [132] Huycke MM, Moore D, Joyce W, Wise P, Shepard L, Kotake Y, *et al*. Extracellular superoxide production by *Enterococcus faecalis* requires demethylmenaquinone and is attenuated by functional terminal quinol oxidases. *Mol Microbiol* 2001;42(3):729–740. doi:10.1046/j.1365-2958.2001.02638.x, PMID:11722738.
- [133] Evans MD, Dizdaroglu M, Cooke MS. Oxidative DNA damage and disease: induction, repair and significance. *Mutat Res* 2004;567(1):1–61. doi:10.1016/j.mrrev.2003.11.001, PMID:15341901.
- [134] Veziant J, Gagnière J, Jouberton E, Bonnin V, Sauvanet P, Pezet D, *et al*. Association of colorectal cancer with pathogenic *Escherichia coli*: Focus on mechanisms using optical imaging. *World J Clin Oncol* 2016;7(3):293–301. doi:10.5306/wjco.v7.i3.293, PMID:27298769.
- [135] Wassenaar TM. *E. coli* and colorectal cancer: a complex relationship that deserves a critical mindset. *Crit Rev Microbiol* 2018;44(5):619–632. doi:10.1080/1040841X.2018.1481013, PMID:29909724.
- [136] Fung KY, Cosgrove L, Lockett T, Head R, Topping DL. A review of the potential mechanisms for the lowering of colorectal oncogenesis by butyrate. *Br J Nutr* 2012;108(5):820–831. doi:10.1017/S0007114512001948, PMID:22676885.
- [137] Sánchez-Alcoholado L, Ramos-Molina B, Otero A, Laborda-Illanes A, Ordóñez R, Medina JA, *et al*. The Role of the Gut Microbiome in Colorectal Cancer Development and Therapy Response. *Cancers (Basel)* 2020;12(6):E1406. doi:10.3390/cancers12061406, PMID:32486066.
- [138] Huang QY, Yao F, Zhou CR, Huang XY, Wang Q, Long H, *et al*. Role of gut microbiome in regulating the effectiveness of metformin in reducing colorectal cancer in type 2 diabetes. *World J Clin Cases* 2020;8(24):6213–6228. doi:10.12998/wjcc.v8.i24.6213, PMID:33392303.
- [139] Bai B, Chen H. Metformin: A Novel Weapon Against Inflammation. *Front Pharmacol* 2021;12:622262. doi:10.3389/fphar.2021.622262, PMID:33584319.
- [140] Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. *Nat Rev Microbiol* 2021;19(1):55–71. doi:10.1038/s41579-020-0433-9, PMID:32887946.
- [141] Borges-Canha M, Portela-Cidade JP, Dinis-Ribeiro M, Leite-Moreira AF, Pimentel-Nunes P. Role of colonic microbiota in colorectal carcinogenesis: a systematic review. *Rev Esp Enferm Dig* 2015;107(11):659–671. doi:10.17235/reed.2015.3830/2015, PMID:26541655.
- [142] Ke H, Li F, Deng W, Li Z, Wang S, Lv P, *et al*. Metformin Exerts Anti-inflammatory and Mucus Barrier Protective Effects by Enriching Akkermansia muciniphila in Mice With Ulcerative Colitis. *Front Pharmacol* 2021;12:726707. doi:10.3389/fphar.2021.726707, PMID:34658866.
- [143] Chambers ES, Preston T, Frost G, Morrison DJ. Role of Gut Microbiota-Generated Short-Chain Fatty Acids in Metabolic and Cardiovascular Health. *Curr Nutr Rep* 2018;7(4):198–206. doi:10.1007/s13668-018-0248-8, PMID:30264354.
- [144] Asadollahi P, Ghanavati R, Rohani M, Razavi S, Esghaei M, Talebi M. Correction: Anti-cancer effects of *Bifidobacterium* species in colon cancer cells and a mouse model of carcinogenesis. *PLoS One* 2020;15(6):e0234777. doi:10.1371/journal.pone.0234777, PMID:32520980.
- [145] Bayerdörffer E, Mannes GA, Richter WO, Ochsenkühn T, Wiebecke B, Köpcke W, *et al*. Increased serum deoxycholic acid levels in men with colorectal adenomas. *Gastroenterology* 1993;104(1):145–151. doi:10.1016/0016-5085(93)90846-5, PMID:8419237.
- [146] O’Keefe SJ. Diet, microorganisms and their metabolites, and colon cancer. *Nat Rev Gastroenterol Hepatol* 2016;13(12):691–706. doi:10.1038/nrgastro.2016.165, PMID:27848961.
- [147] Saus E, Iraola-Guzmán S, Willis JR, Brunet-Vega A, Gabaldón T. Microbiome and colorectal cancer: Roles in carcinogenesis and clinical potential. *Mol Aspects Med* 2019;69:93–106. doi:10.1016/j.mam.2019.05.001, PMID:31082399.
- [148] Butt J, Epplen M. *Helicobacter pylori* and colorectal cancer-A bacterium going abroad? *PLoS Pathog* 2019;15(8):e1007861. doi:10.1371/journal.ppat.1007861, PMID:31393968.
- [149] Courtois S, Bénéjat L, Izotte J, Mégraud F, Varon C, Lehours P, *et al*. Metformin can inhibit *Helicobacter pylori* growth. *Future Microbiol* 2018;13:1575–1583. doi:10.2217/fmb-2018-0184, PMID:30421627.
- [150] Matsuda EM, Oliveira IP, Bao LB, Manzoni FM, Campos NC, Varella BB, *et al*. Impact of covid-19 on people living with HIV-1: care and prevention indicators at a local and nationwide level, Santo André, Brazil. *Rev Saude Publica* 2022;56:37. doi:10.11606/s1518-8787.2022056004314, PMID:35649084.
- [151] Brenchley JM, Price DA, Schacker TW, Asher TE, Silvestri G, Rao S, *et al*. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nat Med* 2006;12(12):1365–1371. doi:10.1038/nm1511, PMID:17115046.
- [152] Coretti L, Natale A, Cuomo M, Florio E, Keller S, Lembo F, *et al*. The Interplay between Defensins and Microbiota in Crohn’s Disease. *Mediators Inflamm* 2017;2017:8392523. doi:10.1155/2017/8392523, PMID:28246439.
- [153] Meade KG, O’Farrelly C.  $\beta$ -Defensins: Farming the Microbiome for Homeostasis and Health. *Front Immunol* 2018;9:3072. doi:10.3389/fimmu.2018.03072, PMID:30761155.
- [154] Zicari S, Sessa L, Cotugno N, Ruggiero A, Morrocchi E, Concato C, *et al*. Immune Activation, Inflammation, and Non-AIDS Co-Morbidities in HIV-Infected Patients under Long-Term ART. *Viruses* 2019;11(3):E200. doi:10.3390/v11030200, PMID:30818749.
- [155] Alzahrani J, Hussain T, Simar D, Palchadhuri R, Abdel-Mohsen M, Crowe SM, *et al*. Inflammatory and immunometabolic consequences of gut dysfunction in HIV: Parallels with IBD and implications for reservoir persistence and non-AIDS comorbidities. *EBioMedicine* 2019;46:522–531. doi:10.1016/j.ebiom.2019.07.027, PMID:31327693.
- [156] Hsu DC, Sereti I. Serious Non-AIDS Events: Therapeutic Targets of Immune Activation and Chronic Inflammation in HIV Infection. *Drugs* 2016;76(5):533–549. doi:10.1007/s40265-016-0546-7, PMID:26915027.
- [157] Vujkovic-Cvijin I, Sortino O, Verheij E, Sklar J, Wit FW, Kootstra NA, *et al*. HIV-associated gut dysbiosis is independent of sexual practice and correlates with noncommunicable diseases. *Nat Commun* 2020;11(1):2448. doi:10.1038/s41467-020-16222-8, PMID:32415070.
- [158] Brenchley JM, Douek DC. HIV infection and the gastrointestinal immune system. *Mucosal Immunol* 2008;1(1):23–30. doi:10.1038/mi.2007.1, PMID:19079157.
- [159] Mehandru S, Tenner-Racz K, Racz P, Markowitz M. The gastrointestinal tract is critical to the pathogenesis of acute HIV-1 infection. *J Allergy Clin Immunol* 2005;116(2):419–422. doi:10.1016/j.jaci.2005.05.040, PMID:16083799.
- [160] Vujkovic-Cvijin I, Dunham RM, Iwai S, Maher MC, Albright RG, Broadhurst MJ, *et al*. Dysbiosis of the gut microbiota is associated with HIV disease progression and tryptophan catabolism. *Sci Transl Med* 2013;5(193):193ra91. doi:10.1126/scitranslmed.3006438, PMID:23843452.
- [161] Vujkovic-Cvijin I, Somsouk M. HIV and the Gut Microbiota: Composition, Consequences, and Avenues for Amelioration. *Curr HIV/AIDS Rep* 2019;16(3):204–213. doi:10.1007/s11904-019-00441-w, PMID:31037552.
- [162] Mutlu EA, Keshavarzian A, Losurdo J, Swanson G, Siewe B, Forsyth C, *et al*. A compositional look at the human gastrointestinal microbiome and immune activation parameters in HIV infected subjects. *PLoS Pathog* 2014;10(2):e1003829. doi:10.1371/journal.ppat.1003829, PMID:24586144.
- [163] Rocafort M, Noguera-Julian M, Rivera J, Pastor L, Guillén Y, Langhorst J, *et al*. Evolution of the gut microbiome following acute HIV-1 infection. *Microbiome* 2019;7(1):73. doi:10.1186/s40168-019-0687-5, PMID:31078141.
- [164] González-Hernández LA, Ruiz-Briseño MDR, Sánchez-Reyes K, Alvarez-Zavala M, Vega-Magaña N, López-Iñiguez A, *et al*. Alterations in bacterial communities, SCFA and biomarkers in an elderly HIV-positive and HIV-negative population in western Mexico. *BMC Infect Dis* 2019;19(1):234. doi:10.1186/s12879-019-3867-9, PMID:30845929.
- [165] Dillon SM, Kibbie J, Lee EJ, Guo K, Santiago ML, Austin GL, *et al*. Low abundance of colonic butyrate-producing bacteria in HIV infection is associated with microbial translocation and immune activation. *AIDS* 2017;31(4):511–521. doi:10.1097/QAD.0000000000001366, PMID:28002063.
- [166] Jin Q, Cheng J, Liu Y, Wu J, Wang X, Wei S, *et al*. Improvement of



- functional recovery by chronic metformin treatment is associated with enhanced alternative activation of microglia/macrophages and increased angiogenesis and neurogenesis following experimental stroke. *Brain Behav Immun* 2014;40:131–142. doi:10.1016/j.bbi.2014.03.003, PMID:24632338.
- [167] Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, *et al*. MetaHIT consortium. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature* 2015;528(7581):262–266. doi:10.1038/nature15766, PMID:26633628.
- [168] Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, *et al*. MetaHIT consortium. Corrigendum: Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature* 2017;545(7652):116. doi:10.1038/nature22318, PMID:28470190.
- [169] Ouyang J, Isnard S, Lin J, Fombuena B, Murette A, Routy B, *et al*. Metformin effect on gut microbiota: insights for HIV-related inflammation. *AIDS Res Ther* 2020;17(1):10. doi:10.1186/s12981-020-00267-2, PMID:32156291.
- [170] Isnard S, Lin J, Fombuena B, Ouyang J, Varin TV, Richard C, *et al*. Repurposing Metformin in Nondiabetic People With HIV: Influence on Weight and Gut Microbiota. *Open Forum Infect Dis* 2020;7(9):ofaa338. doi:10.1093/ofid/ofaa338, PMID:32964062.
- [171] Mendis S, Graham I, Narula J. Addressing the Global Burden of Cardiovascular Diseases; Need for Scalable and Sustainable Frameworks. *Glob Heart* 2022;17(1):48. doi:10.5334/gh.1139, PMID:36051329.
- [172] Whiting DR, Guariguata L, Weil C, Shaw J. IDF diabetes atlas: global estimates of the prevalence of diabetes for 2011 and 2030. *Diabetes Res Clin Pract* 2011;94(3):311–321. doi:10.1016/j.diabres.2011.10.029, PMID:22079683.
- [173] Loh M, Zhou L, Ng HK, Chambers JC. Epigenetic disturbances in obesity and diabetes: Epidemiological and functional insights. *Mol Metab* 2019;27S:S33–S41. doi:10.1016/j.molmet.2019.06.011, PMID:3150829.
- [174] Barr EL, Zimmet PZ, Welborn TA, Jolley D, Magliano DJ, Dunstan DW, *et al*. Risk of cardiovascular and all-cause mortality in individuals with diabetes mellitus, impaired fasting glucose, and impaired glucose tolerance: the Australian Diabetes, Obesity, and Lifestyle Study (AusDiab). *Circulation* 2007;116(2):151–157. doi:10.1161/CIRCULATIONAHA.106.685628, PMID:17576864.
- [175] Zhou W, Cheng Y, Zhu P, Nasser MI, Zhang X, Zhao M. Implication of Gut Microbiota in Cardiovascular Diseases. *Oxid Med Cell Longev* 2020;2020:5394096. doi:10.1155/2020/5394096, PMID:33062141.
- [176] Mozaffarian D, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, Cushman M, *et al*. Writing Group Members., American Heart Association Statistics Committee., Stroke Statistics Subcommittee. Executive Summary: Heart Disease and Stroke Statistics—2016 Update: A Report From the American Heart Association. *Circulation* 2016;133(4):447–454. doi:10.1161/CIR.0000000000000366, PMID:26811276.
- [177] Ripatti S, Tikkanen E, Orho-Melander M, Havulinna AS, Silander K, Sharma A, *et al*. A multilocus genetic risk score for coronary heart disease: case-control and prospective cohort analyses. *Lancet* 2010;376(9750):1393–1400. doi:10.1016/S0140-6736(10)61267-6, PMID:20971364.
- [178] Brown JM, Hazen SL. Microbial modulation of cardiovascular disease. *Nat Rev Microbiol* 2018;16(3):171–181. doi:10.1038/nrmicro.2017.149, PMID:29307889.
- [179] Tang WH, Kitai T, Hazen SL. Gut Microbiota in Cardiovascular Health and Disease. *Circ Res* 2017;120(7):1183–1196. doi:10.1161/CIRCRESAHA.117.309715, PMID:28360349.
- [180] Gill SR, Pop M, Deboy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, *et al*. Metagenomic analysis of the human distal gut microbiome. *Science* 2006;312(5778):1355–1359. doi:10.1126/science.1124234, PMID:16741115.
- [181] Emoto T, Yamashita T, Sasaki N, Hirota Y, Hayashi T, So A, *et al*. Analysis of Gut Microbiota in Coronary Artery Disease Patients: a Possible Link between Gut Microbiota and Coronary Artery Disease. *J Atheroscler Thromb* 2016;23(8):908–921. doi:10.5551/jat.32672, PMID:26947598.
- [182] Jie Z, Xia H, Zhong SL, Feng Q, Li S, Liang S, *et al*. The gut microbiome in atherosclerotic cardiovascular disease. *Nat Commun* 2017;8(1):845. doi:10.1038/s41467-017-00900-1, PMID:29018189.
- [183] Zhu Q, Gao R, Zhang Y, Pan D, Zhu Y, Zhang X, *et al*. Dysbiosis signatures of gut microbiota in coronary artery disease. *Physiol Genomics* 2018;50(10):893–903. doi:10.1152/physiolgenomics.00070.2018, PMID:30192713.
- [184] Neves AL, Coelho J, Couto L, Leite-Moreira A, Roncon-Albuquerque R Jr. Metabolic endotoxemia: a molecular link between obesity and cardiovascular risk. *J Mol Endocrinol* 2013;51(2):R51–R64. doi:10.1530/JME-13-0079, PMID:23943858.
- [185] Medzhitov R. Recognition of microorganisms and activation of the immune response. *Nature* 2007;449(7164):819–826. doi:10.1038/nature06246, PMID:17943118.
- [186] Heidenreich PA, Trogon JG, Khavjou OA, Butler J, Dracup K, Ezekowitz MD, *et al*. Forecasting the future of cardiovascular disease in the United States: a policy statement from the American Heart Association. *Circulation* 2011;123(8):933–944. doi:10.1161/CIR.Ob013e31820a55f5, PMID:21262990.
- [187] Tang WH, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, *et al*. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med* 2013;368(17):1575–1584. doi:10.1056/NEJMoa1109400, PMID:23614584.
- [188] Liu Y, Dai M. Trimethylamine N-Oxide Generated by the Gut Microbiota Is Associated with Vascular Inflammation: New Insights into Atherosclerosis. *Mediators Inflamm* 2020;2020:4634172. doi:10.1155/2020/4634172, PMID:32148438.
- [189] Schiattarella GG, Sannino A, Toscano E, Giugliano G, Gargiulo G, Franzone A, *et al*. Gut microbe-generated metabolite trimethylamine-N-oxide as cardiovascular risk biomarker: a systematic review and dose-response meta-analysis. *Eur Heart J* 2017;38(39):2948–2956. doi:10.1093/eurheartj/ehx342, PMID:29020409.
- [190] Bennett BJ, de Aguiar Vallim TQ, Wang Z, Shih DM, Meng Y, Gregory J, *et al*. Trimethylamine-N-oxide, a metabolite associated with atherosclerosis, exhibits complex genetic and dietary regulation. *Cell Metab* 2013;17(1):49–60. doi:10.1016/j.cmet.2012.12.011, PMID:23312283.
- [191] Yang S, Li X, Yang F, Zhao R, Pan X, Liang J, *et al*. Gut Microbiota-Dependent Marker TMAO in Promoting Cardiovascular Disease: Inflammation Mechanism, Clinical Prognostic, and Potential as a Therapeutic Target. *Front Pharmacol* 2019;10:1360. doi:10.3389/fphar.2019.01360, PMID:31803054.
- [192] Mayerhofer CCK, Ueland T, Broch K, Vincent RP, Cross GF, Dahl CP, *et al*. Increased Secondary/Primary Bile Acid Ratio in Chronic Heart Failure. *J Card Fail* 2017;23(9):666–671. doi:10.1016/j.cardfail.2017.06.007, PMID:28688889.
- [193] Jones ML, Martoni CJ, Parent M, Prakash S. Cholesterol-lowering efficacy of a microencapsulated bile salt hydrolase-active *Lactobacillus reuteri* NCIMB 30242 yoghurt formulation in hypercholesterolemic adults. *Br J Nutr* 2012;107(10):1505–1513. doi:10.1017/S0007114511004703, PMID:22067612.
- [194] Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. *Nature* 2012;489(7415):242–249. doi:10.1038/nature11552, PMID:22972297.
- [195] Ridlon JM, Harris SC, Bhowmik S, Kang DJ, Hylemon PB. Consequences of bile salt biotransformations by intestinal bacteria. *Gut Microbes* 2016;7(1):22–39. doi:10.1080/19490976.2015.1127483, PMID:26939849.
- [196] Chiang JY. Bile acids: regulation of synthesis. *J Lipid Res* 2009;50(10):1955–1966. doi:10.1194/jlr.R900010-JLR200, PMID:19346330.
- [197] Hansson GK, Robertson AK, Söderberg-Nauclér C. Inflammation and atherosclerosis. *Annu Rev Pathol* 2006;1:297–329. doi:10.1146/annurev.pathol.1.110304.100100, PMID:18039117.
- [198] Joyce SA, Gahan CG. Disease-Associated Changes in Bile Acid Profiles and Links to Altered Gut Microbiota. *Dig Dis* 2017;35(3):169–177. doi:10.1159/000450907, PMID:28249284.
- [199] Yang R, Dong J, Zhao H, Li H, Guo H, Wang S, *et al*. Association of branched-chain amino acids with carotid intima-media thickness and coronary artery disease risk factors. *PLoS One* 2014;9(6):e99598. doi:10.1371/journal.pone.0099598, PMID:24910999.
- [200] Yang P, Hu W, Fu Z, Sun L, Zhou Y, Gong Y, *et al*. The positive association of branched-chain amino acids and metabolic dyslipidemia in Chinese

- Han population. *Lipids Health Dis* 2016;15:120. doi:10.1186/s12944-016-0291-7, PMID:27457614.
- [201] Mels CM, Schutte AE, Schutte R, Huisman HW, Smith W, Fourie CM, *et al*. The link between vascular deterioration and branched chain amino acids in a population with high glycated haemoglobin: the SABPA study. *Amino Acids* 2013;45(6):1405–1413. doi:10.1007/s00726-013-1611-0, PMID:24178767.
- [202] Magnusson M, Lewis GD, Ericson U, Orho-Melander M, Hedblad B, Engström G, *et al*. A diabetes-predictive amino acid score and future cardiovascular disease. *Eur Heart J* 2013;34(26):1982–1989. doi:10.1093/eurheartj/ehs424, PMID:23242195.
- [203] Tobias DK, Mora S, Verma S, Lawler PR. Altered branched chain amino acid metabolism: toward a unifying cardiometabolic hypothesis. *Curr Opin Cardiol* 2018;33(5):558–564. doi:10.1097/HCO.0000000000000552, PMID:29994805.
- [204] van Stee MF, de Graaf AA, Groen AK. Actions of metformin and statins on lipid and glucose metabolism and possible benefit of combination therapy. *Cardiovasc Diabetol* 2018;17(1):94. doi:10.1186/s12933-018-0738-4, PMID:29960584.
- [205] Monami M, Zannoni S, Pala L, Silverii A, Andreozzi F, Sesti G, *et al*. Effects of glucagon-like peptide-1 receptor agonists on mortality and cardiovascular events: A comprehensive meta-analysis of randomized controlled trials. *Int J Cardiol* 2017;240:414–421. doi:10.1016/j.ijcard.2017.03.163, PMID:28501352.
- [206] Ding QY, Tian JX, Li M, Lian FM, Zhao LH, Wei XX, *et al*. Interactions Between Therapeutics for Metabolic Disease, Cardiovascular Risk Factors, and Gut Microbiota. *Front Cell Infect Microbiol* 2020;10:530160. doi:10.3389/fcimb.2020.530160, PMID:33194785.
- [207] Wu H, Esteve E, Tremaroli V, Khan MT, Caesar R, Mannerås-Holm L, *et al*. Metformin alters the gut microbiome of individuals with treatment-naïve type 2 diabetes, contributing to the therapeutic effects of the drug. *Nat Med* 2017;23(7):850–858. doi:10.1038/nm.4345, PMID:28530702.
- [208] Plovier H, Everard A, Druart C, Depommier C, Van Hul M, Geurts L, *et al*. A purified membrane protein from *Akkermansia muciniphila* or the pasteurized bacterium improves metabolism in obese and diabetic mice. *Nat Med* 2017;23(1):107–113. doi:10.1038/nm.4236, PMID:27892954.
- [209] Chelakkot C, Choi Y, Kim DK, Park HT, Ghim J, Kwon Y, *et al*. *Akkermansia muciniphila*-derived extracellular vesicles influence gut permeability through the regulation of tight junctions. *Exp Mol Med* 2018;50(2):e450. doi:10.1038/emmm.2017.282, PMID:29472701.
- [210] Matsubara T, Li F, Gonzalez FJ. FXR signaling in the enterohepatic system. *Mol Cell Endocrinol* 2013;368(1-2):17–29. doi:10.1016/j.mce.2012.05.004, PMID:22609541.
- [211] Zhang Y, Wang X, Vales C, Lee FY, Lee H, Lusis AJ, *et al*. FXR deficiency causes reduced atherosclerosis in *Ldlr*<sup>-/-</sup> mice. *Arterioscler Thromb Vasc Biol* 2006;26(10):2316–2321. doi:10.1161/01.ATV.0000235697.35431.05, PMID:16825595.
- [212] Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, *et al*. Gut metagenome in European women with normal, impaired and diabetic glucose control. *Nature* 2013;498(7452):99–103. doi:10.1038/nature12198, PMID:23719380.
- [213] Fujisaka S, Avila-Pacheco J, Soto M, Kostic A, Dreyfuss JM, Pan H, *et al*. Diet, Genetics, and the Gut Microbiome Drive Dynamic Changes in Plasma Metabolites. *Cell Rep* 2018;22(11):3072–3086. doi:10.1016/j.celrep.2018.02.060, PMID:29539432.
- [214] Kuka J, Videja M, Makrecka-Kuka M, Liepins J, Grinberga S, Sevostjanovs E, *et al*. Metformin decreases bacterial trimethylamine production and trimethylamine N-oxide levels in db/db mice. *Sci Rep* 2020;10(1):14555. doi:10.1038/s41598-020-71470-4, PMID:32884086.
- [215] Su C, Li X, Yang Y, Du Y, Zhang X, Wang L, *et al*. Metformin alleviates choline diet-induced TMAO elevation in C57BL/6J mice by influencing gut-microbiota composition and functionality. *Nutr Diabetes* 2021;11(1):27. doi:10.1038/s41387-021-00169-w, PMID:34389700.
- [216] Livingston G, Huntley J, Sommerlad A, Ames D, Ballard C, Banerjee S, *et al*. Dementia prevention, intervention, and care: 2020 report of the Lancet Commission. *Lancet* 2020;396(10248):413–446. doi:10.1016/S0140-6736(20)30367-6, PMID:32738937.
- [217] Alzheimer's Association. Alzheimer's disease facts and figures includes a special report on the financial and personal benefits of early diagnosis. Chicago: Alzheimer's Association; 2018.
- [218] Scheltens P, De Strooper B, Kivipelto M, Holstege H, Chételat G, Teunissen CE, *et al*. Alzheimer's disease. *Lancet* 2021;397(10284):1577–1590. doi:10.1016/S0140-6736(20)32205-4, PMID:33667416.
- [219] Wu S, Liu X, Jiang R, Yan X, Ling Z. Roles and Mechanisms of Gut Microbiota in Patients With Alzheimer's Disease. *Front Aging Neurosci* 2021;13:650047. doi:10.3389/fnagi.2021.650047, PMID:34122039.
- [220] Guo T, Zhang D, Zeng Y, Huang TY, Xu H, Zhao Y. Molecular and cellular mechanisms underlying the pathogenesis of Alzheimer's disease. *Mol Neurodegener* 2020;15(1):40. doi:10.1186/s13024-020-00391-7, PMID:32677986.
- [221] Zhang M, Zhao D, Zhou G, Li C. Dietary Pattern, Gut Microbiota, and Alzheimer's Disease. *J Agric Food Chem* 2020;68(46):12800–12809. doi:10.1021/acs.jafc.9b08309, PMID:32090565.
- [222] Zolochovska O, Tagliatalata G. Selected microRNAs Increase Synaptic Resilience to the Damaging Binding of the Alzheimer's Disease Amyloid Beta Oligomers. *Mol Neurobiol* 2020;57(5):2232–2243. doi:10.1007/s12035-020-01868-8, PMID:31997075.
- [223] Ashford MT, Veitch DP, Neuhaus J, Nosheny RL, Tosun D, Weiner MW. The search for a convenient procedure to detect one of the earliest signs of Alzheimer's disease: A systematic review of the prediction of brain amyloid status. *Alzheimers Dement* 2021;17(5):866–887. doi:10.1002/alz.12253, PMID:33583100.
- [224] Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002;297(5580):353–356. doi:10.1126/science.1072994, PMID:12130773.
- [225] Goedert M. Tau filaments in neurodegenerative diseases. *FEBS Lett* 2018;592(14):2383–2391. doi:10.1002/1873-3468.13108, PMID:29790176.
- [226] Cenit MC, Nuevo IC, Codoñer-Franch P, Dinan TG, Sanz Y. Gut microbiota and attention deficit hyperactivity disorder: new perspectives for a challenging condition. *Eur Child Adolesc Psychiatry* 2017;26(9):1081–1092. doi:10.1007/s00787-017-0969-z, PMID:28289903.
- [227] Caini S, Bagnoli S, Palli D, Saieva C, Ceroti M, Bendinelli B, *et al*. Total and cancer mortality in a cohort of ulcerative colitis and Crohn's disease patients: The Florence inflammatory bowel disease study, 1978-2010. *Dig Liver Dis* 2016;48(10):1162–1167. doi:10.1016/j.dld.2016.07.008, PMID:27481588.
- [228] Chen CH, Lin CL, Kao CH. Irritable Bowel Syndrome Is Associated with an Increased Risk of Dementia: A Nationwide Population-Based Study. *PLoS One* 2016;11(1):e0144589. doi:10.1371/journal.pone.0144589, PMID:26731277.
- [229] Cattaneo A, Cattaneo N, Galluzzi S, Provasi S, Lopizzo N, Festari C, *et al*. INDIA-FBP Group. Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. *Neurobiol Aging* 2017;49:60–68. doi:10.1016/j.neurobiolaging.2016.08.019, PMID:27776263.
- [230] Li B, He Y, Ma J, Huang P, Du J, Cao L, *et al*. Mild cognitive impairment has similar alterations as Alzheimer's disease in gut microbiota. *Alzheimers Dement* 2019;15(10):1357–1366. doi:10.1016/j.jalz.2019.07.002, PMID:31434623.
- [231] Diaz Heijtz R, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A, *et al*. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci U S A* 2011;108(7):3047–3052. doi:10.1073/pnas.1010529108, PMID:21282636.
- [232] Desbonnet L, Clarke G, Traplin A, O'Sullivan O, Crispie F, Moloney RD, *et al*. Gut microbiota depletion from early adolescence in mice: Implications for brain and behaviour. *Brain Behav Immun* 2015;48:165–173. doi:10.1016/j.bbi.2015.04.004, PMID:25866195.
- [233] Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat Rev Neurosci* 2012;13(10):701–712. doi:10.1038/nrn3346, PMID:22968153.
- [234] Burokas A, Moloney RD, Dinan TG, Cryan JF. Microbiota regulation of the Mammalian gut-brain axis. *Adv Appl Microbiol* 2015;91:1–62. doi:10.1016/bs.aambs.2015.02.001, PMID:25911232.
- [235] Morris G, Berk M, Carvalho A, Caso JR, Sanz Y, Walder K, *et al*. The Role of the Microbial Metabolites Including Tryptophan Catabolites and Short Chain Fatty Acids in the Pathophysiology of Immune-Inflammatory and Neuroimmune Disease. *Mol Neurobiol* 2017;54(6):4432–4451. doi:10.1007/s12035-016-0004-2, PMID:27349436.

- [236] Oli MW, Otoo HN, Crowley PJ, Heim KP, Nascimento MM, Ramscook CB, *et al*. Functional amyloid formation by *Streptococcus mutans*. *Microbiology (Reading)* 2012;158(Pt 12):2903–2916. doi:10.1099/mic.0.060855-0, PMID:23082034.
- [237] Cheng G, Huang C, Deng H, Wang H. Diabetes as a risk factor for dementia and mild cognitive impairment: a meta-analysis of longitudinal studies. *Intern Med J* 2012;42(5):484–491. doi:10.1111/j.1445-5994.2012.02758.x, PMID:22372522.
- [238] Pistollato F, Sumalla Cano S, Elio I, Masias Vergara M, Giampieri F, Battino M. Role of gut microbiota and nutrients in amyloid formation and pathogenesis of Alzheimer disease. *Nutr Rev* 2016;74(10):624–634. doi:10.1093/nutrit/nuw023, PMID:27634977.
- [239] Paranjayee N, Daggett V. De Novo Designed  $\alpha$ -Sheet Peptides Inhibit Functional Amyloid Formation of *Streptococcus mutans* Biofilms. *J Mol Biol* 2018;430(20):3764–3773. doi:10.1016/j.jmb.2018.07.005, PMID:30006266.
- [240] Vogt NM, Romano KA, Darst BF, Engelman CD, Johnson SC, Carlsson CM, *et al*. The gut microbiota-derived metabolite trimethylamine N-oxide is elevated in Alzheimer's disease. *Alzheimers Res Ther* 2018;10(1):124. doi:10.1186/s13195-018-0451-2, PMID:30579367.
- [241] Xiao Y, Liu F, Li S, Jiang N, Yu C, Zhu X, *et al*. Metformin promotes innate immunity through a conserved PMK-1/p38 MAPK pathway. *Virulence* 2020;11(1):39–48. doi:10.1080/21505594.2019.1706305, PMID:31851866.
- [242] Böhme J, Martinez N, Li S, Lee A, Marzuki M, Tizazu AM, *et al*. Metformin enhances anti-mycobacterial responses by educating CD8+ T-cell immunometabolic circuits. *Nat Commun* 2020;11(1):5225. doi:10.1038/s41467-020-19095-z, PMID:33067434.
- [243] Chen Y, Zhao S, Fan Z, Li Z, Zhu Y, Shen T, *et al*. Metformin attenuates plaque-associated tau pathology and reduces amyloid- $\beta$  burden in APP/PS1 mice. *Alzheimers Res Ther* 2021;13(1):40. doi:10.1186/s13195-020-00761-9, PMID:33563332.
- [244] Bruce-Keller AJ, Salbaum JM, Luo M, Blanchard E IV, Taylor CM, Welsh DA, *et al*. Obese-type gut microbiota induce neurobehavioral changes in the absence of obesity. *Biol Psychiatry* 2015;77(7):607–615. doi:10.1016/j.biopsych.2014.07.012, PMID:25173628.
- [245] Ma X, Xiao W, Li H, Pang P, Xue F, Wan L, *et al*. Metformin restores hippocampal neurogenesis and learning and memory via regulating gut microbiota in the obese mouse model. *Brain Behav Immun* 2021;95:68–83. doi:10.1016/j.bbi.2021.02.011, PMID:33609653.
- [246] Pugazhenthis S, Qin L, Reddy PH. Common neurodegenerative pathways in obesity, diabetes, and Alzheimer's disease. *Biochim Biophys Acta Mol Basis Dis* 2017;1863(5):1037–1045. doi:10.1016/j.bbadis.2016.04.017, PMID:27156888.
- [247] Xu J, Liang R, Zhang W, Tian K, Li J, Chen X, *et al*. Faecalibacterium prausnitzii-derived microbial anti-inflammatory molecule regulates intestinal integrity in diabetes mellitus mice via modulating tight junction protein expression. *J Diabetes* 2020;12(3):224–236. doi:10.1111/1753-0407.12986, PMID:31503404.
- [248] Koike K, Moore EE, Moore FA, Read RA, Carl VS, Banerjee A. Gut ischemia/reperfusion produces lung injury independent of endotoxin. *Crit Care Med* 1994;22(9):1438–1444. doi:10.1097/00003246-199409000-00014, PMID:8062567.
- [249] de Groot H, Rauhen U. Ischemia-reperfusion injury: processes in pathogenetic networks: a review. *Transplant Proc* 2007;39(2):481–484. doi:10.1016/j.transproceed.2006.12.012, PMID:17362763.
- [250] Jia Y, Cui R, Wang C, Feng Y, Li Z, Tong Y, *et al*. Metformin protects against intestinal ischemia-reperfusion injury and cell pyroptosis via TXNIP-NLRP3-GSDMD pathway. *Redox Biol* 2020;32:101534. doi:10.1016/j.redox.2020.101534, PMID:32330868.
- [251] Rogina B, Reenan RA, Nilsen SP, Helfand SL. Extended life-span conferred by cotransporter gene mutations in *Drosophila*. *Science* 2000;290(5499):2137–2140. doi:10.1126/science.290.5499.2137, PMID:1118146.
- [252] Officer A, Thiyagarajan JA, Schneiders ML, Nash P, de la Fuente-Núñez V. Ageism, Healthy Life Expectancy and Population Ageing: How Are They Related? *Int J Environ Res Public Health* 2020;17(9):3159. doi:10.3390/ijerph17093159, PMID:32370093.
- [253] López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell* 2013;153(6):1194–1217. doi:10.1016/j.cell.2013.05.039, PMID:23746838.
- [254] Bana B, Cabreiro F. The Microbiome and Aging. *Annu Rev Genet* 2019;53:239–261. doi:10.1146/annurev-genet-112618-043650, PMID:31487470.
- [255] Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, *et al*. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci* 2000;908:244–254. doi:10.1111/j.1749-6632.2000.tb06651.x, PMID:10911963.
- [256] Ferrucci L, Fabbri E. Inflammageing: chronic inflammation in ageing, cardiovascular disease, and frailty. *Nat Rev Cardiol* 2018;15(9):505–522. doi:10.1038/s41569-018-0064-2, PMID:30065258.
- [257] Gorelick PB. Role of inflammation in cognitive impairment: results of observational epidemiological studies and clinical trials. *Ann N Y Acad Sci* 2010;1207:155–162. doi:10.1111/j.1749-6632.2010.05726.x, PMID:20955439.
- [258] Bektas A, Schurman SH, Sen R, Ferrucci L. Aging, inflammation and the environment. *Exp Gerontol* 2018;105:10–18. doi:10.1016/j.exger.2017.12.015, PMID:29275161.
- [259] Simpson RJ. Aging and inflammation: Directing traffic through physical activity. *Brain Behav Immun* 2016;56:10–11. doi:10.1016/j.bbi.2016.05.015, PMID:27223097.
- [260] Wu L, Zeng T, Zinellu A, Rubino S, Kelvin DJ, Carru C. A Cross-Sectional Study of Compositional and Functional Profiles of Gut Microbiota in Sardinian Centenarians. *mSystems* 2019;4(4):e00325–19. doi:10.1128/mSystems.00325-19, PMID:31289141.
- [261] Kushugulova A, Kozhakhmetov S, Baishanova D, Tynybayeva I, Kakiyeva A, Khassenbekova Z, *et al*. Gut microbiome diversity in kazakhstani women of different age groups. *Int J Probiotics Prebiotics* 2015;10(2-3):97–108.
- [262] Drago L, Toscano M, Rodighiero V, De Vecchi E, Mogna G. Cultivable and pyrosequenced fecal microflora in centenarians and young subjects. *J Clin Gastroenterol* 2012;46 Suppl:S81–S84. doi:10.1097/MCG.0b013e3182693982, PMID:22955365.
- [263] Schlender L, Martinez YV, Adeniji C, Reeves D, Faller B, Sommerauer C, *et al*. Efficacy and safety of metformin in the management of type 2 diabetes mellitus in older adults: a systematic review for the development of recommendations to reduce potentially inappropriate prescribing. *BMC Geriatr* 2017;17(Suppl 1):227. doi:10.1186/s12877-017-0574-5, PMID:29047344.
- [264] Vallianou N, Stratigou T, Christodoulatos GS, Dalamaga M. Understanding the Role of the Gut Microbiome and Microbial Metabolites in Obesity and Obesity-Associated Metabolic Disorders: Current Evidence and Perspectives. *Curr Obes Rep* 2019;8(3):317–332. doi:10.1007/s13679-019-00352-2, PMID:31175629.
- [265] Hong J, Zhang Y, Lai S, Lv A, Su Q, Dong Y, *et al*. SPREAD-DIMCAD Investigators. Effects of metformin versus glipizide on cardiovascular outcomes in patients with type 2 diabetes and coronary artery disease. *Diabetes Care* 2013;36(5):1304–1311. doi:10.2337/dc12-0719, PMID:23230096.
- [266] Cazzaniga M, DeCensi A, Pruneri G, Puntoni M, Bottiglieri L, Varrichio C, *et al*. The effect of metformin on apoptosis in a breast cancer presurgical trial. *Br J Cancer* 2013;109(11):2792–2797. doi:10.1038/bjc.2013.657, PMID:24157825.
- [267] Anisimov VN. Do metformin a real anticarcinogen? A critical reappraisal of experimental data. *Ann Transl Med* 2014;2(6):60. doi:10.3978/j.issn.2305-5839.2014.06.02, PMID:25333035.
- [268] Mofu Mato EP, Guewo-Fokeng M, Essop MF, Owira PMO. Genetic polymorphisms of organic cation transporter 1 (OCT1) and responses to metformin therapy in individuals with type 2 diabetes: A systematic review. *Medicine (Baltimore)* 2018;97(27):e11349. doi:10.1097/MD.00000000000011349, PMID:29979413.
- [269] Shu Y, Sheardown SA, Brown C, Owen RP, Zhang S, Castro RA, *et al*. Effect of genetic variation in the organic cation transporter 1 (OCT1) on metformin action. *J Clin Invest* 2007;117(5):1422–1431. doi:10.1172/JCI30558, PMID:17476361.
- [270] Sam WJ, Roza O, Hon YY, Alfaro RM, Calis KA, Reynolds JC, *et al*. Effects of SLC22A1 Polymorphisms on Metformin-Induced Reductions in Adiposity and Metformin Pharmacokinetics in Obese Children With Insulin Resistance. *J Clin Pharmacol* 2017;57(2):219–229. doi:10.1002/jcph.796, PMID:27407018.
- [271] Saad MJ, Santos A, Prada PO. Linking Gut Microbiota and Inflammation to Obesity and Insulin Resistance. *Physiology (Bethesda)* 2016;

- 31(4):283–293. doi:10.1152/physiol.00041.2015, PMID:27252163.
- [272] Bulterijs S. Metformin as a geroprotector. *Rejuvenation Res* 2011;14(5):469–482. doi:10.1089/rej.2011.1153, PMID:21882902.
- [273] Berstein LM. Metformin in obesity, cancer and aging: addressing controversies. *Aging (Albany NY)* 2012;4(5):320–329. doi:10.18632/aging.100455, PMID:22589237.
- [274] Miles JM, Rule AD, Borlaug BA. Use of metformin in diseases of aging. *Curr Diab Rep* 2014;14(6):490. doi:10.1007/s11892-014-0490-4, PMID:24752835.
- [275] Eurich DT, Weir DL, Majumdar SR, Tsuyuki RT, Johnson JA, Tjosvold L, *et al*. Comparative safety and effectiveness of metformin in patients with diabetes mellitus and heart failure: systematic review of observational studies involving 34,000 patients. *Circ Heart Fail* 2013;6(3):395–402. doi:10.1161/CIRCHEARTFAILURE.112.000162, PMID:23508758.
- [276] Patrone C, Eriksson O, Lindholm D. Diabetes drugs and neurological disorders: new views and therapeutic possibilities. *Lancet Diabetes Endocrinol* 2014;2(3):256–262. doi:10.1016/S2213-8587(13)70125-6, PMID:24622756.
- [277] Praticchizzo F, Giuliani A, Mensà E, Sabbatinelli J, De Nigris V, Rippon MR, *et al*. Pleiotropic effects of metformin: Shaping the microbiome to manage type 2 diabetes and postpone ageing. *Ageing Res Rev* 2018;48:87–98. doi:10.1016/j.arr.2018.10.003, PMID:30336272.
- [278] Mueller NT, Differding MK, Zhang M, Maruthur NM, Juraschek SP, Miller ER 3rd, *et al*. Metformin Affects Gut Microbiome Composition and Function and Circulating Short-Chain Fatty Acids: A Randomized Trial. *Diabetes Care* 2021;44(7):1462–1471. doi:10.2337/dc20-2257, PMID:34006565.
- [279] Buford TW. Dis)Trust your gut: the gut microbiome in age-related inflammation, health, and disease. *Microbiome* 2017;5(1):80. doi:10.1186/s40168-017-0296-0, PMID:28709450.
- [280] Nagpal R, Mainali R, Ahmadi S, Wang S, Singh R, Kavanagh K, *et al*. Gut microbiome and aging: Physiological and mechanistic insights. *Nutr Healthy Aging* 2018;4(4):267–285. doi:10.3233/NHA-170030, PMID:29951588.
- [281] Ahmadi S, Razazan A, Nagpal R, Jain S, Wang B, Mishra SP, *et al*. Metformin Reduces Aging-Related Leaky Gut and Improves Cognitive Function by Beneficially Modulating Gut Microbiome/Goblet Cell/Mucin Axis. *J Gerontol A Biol Sci Med Sci* 2020;75(7):e9–e21. doi:10.1093/gerona/glaa056, PMID:32129462.
- [282] Salvestrini V, Sell C, Lorenzini A. Obesity May Accelerate the Aging Process. *Front Endocrinol (Lausanne)* 2019;10:266. doi:10.3389/fendo.2019.00266, PMID:31130916.
- [283] Ahima RS. Connecting obesity, aging and diabetes. *Nat Med* 2009;15(9):996–997. doi:10.1038/nm0909-996, PMID:19734871.
- [284] Mackenbach JP, Karanikolos M, Looman CW. The rise of mortality from mental and neurological diseases in Europe, 1979–2009: observational study. *BMC Public Health* 2014;14:840. doi:10.1186/1471-2458-14-840, PMID:25118099.
- [285] Apostu SA, Vasile V, Sava V. Do Cardiovascular Diseases Significantly Influence Healthy Aging? *Int J Environ Res Public Health* 2021;18(14):7226. doi:10.3389/ijerph18147226, PMID:34299677.
- [286] Botta L, Dal Maso L, Guzzinati S, Panato C, Gatta G, Trama A, *et al*. Changes in life expectancy for cancer patients over time since diagnosis. *J Adv Res* 2019;20:153–159. doi:10.1016/j.jare.2019.07.002, PMID:31467707.
- [287] Mohammed I, Hollenberg MD, Ding H, Triggler CR. A Critical Review of the Evidence That Metformin Is a Putative Anti-Aging Drug That Enhances Healthspan and Extends Lifespan. *Front Endocrinol (Lausanne)* 2021;12:718942. doi:10.3389/fendo.2021.718942, PMID:34421827.
- [288] Jensen JB, Sundelin EI, Jakobsen S, Gormsen LC, Munk OL, Frøkiær J, *et al*. [<sup>11</sup>C]-Labeled Metformin Distribution in the Liver and Small Intestine Using Dynamic Positron Emission Tomography in Mice Demonstrates Tissue-Specific Transporter Dependency. *Diabetes* 2016;65(6):1724–1730. doi:10.2337/db16-0032, PMID:26993065.
- [289] Han TK, Proctor WR, Costales CL, Cai H, Everett RS, Thakker DR. Four cation-selective transporters contribute to apical uptake and accumulation of metformin in Caco-2 cell monolayers. *J Pharmacol Exp Ther* 2015;352(3):519–528. doi:10.1124/jpet.114.220350, PMID:25563903.
- [290] Mostafa-Hedeab G, Mohamed AA, Ebid GT, Sabry D, Salam RF, Has-sen ME. Effect of MATE 1, MATE 2 and OCT1 single nucleotide polymorphisms on metformin action in recently diagnosed Egyptian type-2 diabetic patients. *Biomed Pharmacol J* 2018;11(1):149–157. doi:10.13005/bpj/1356.
- [291] Wang DS, Jonker JW, Kato Y, Kusu-hara H, Schinkel AH, Sugiyama Y. Involvement of organic cation transporter 1 in hepatic and intestinal distribution of metformin. *J Pharmacol Exp Ther* 2002;302(2):510–515. doi:10.1124/jpet.102.034140, PMID:12130709.
- [292] Sambol NC, Chiang J, O’Conner M, Liu CY, Lin ET, Goodman AM, *et al*. Pharmacokinetics and pharmacodynamics of metformin in healthy subjects and patients with noninsulin-dependent diabetes mellitus. *J Clin Pharmacol* 1996;36(11):1012–1021. doi:10.1177/009127009603601105, PMID:8973990.
- [293] Pentikäinen PJ, Neuvonen PJ, Penttilä A. Pharmacokinetics of metformin after intravenous and oral administration to man. *Eur J Clin Pharmacol* 1979;16(3):195–202. doi:10.1007/BF00562061, PMID:499320.
- [294] Tucker GT, Casey C, Phillips PJ, Connor H, Ward JD, Woods HF. Metformin kinetics in healthy subjects and in patients with diabetes mellitus. *Br J Clin Pharmacol* 1981;12(2):235–246. doi:10.1111/j.1365-2125.1981.tb01206.x, PMID:7306436.
- [295] Zolk O. Disposition of metformin: variability due to polymorphisms of organic cation transporters. *Ann Med* 2012;44(2):119–129. doi:10.3109/07853890.2010.549144, PMID:21366511.
- [296] Takane H, Shikata E, Otsubo K, Higuchi S, Ieiri I. Polymorphism in human organic cation transporters and metformin action. *Pharmacogenomics* 2008;9(4):415–422. doi:10.2217/14622416.9.4.415, PMID:18384255.
- [297] Seitz T, Stalmann R, Dalila N, Chen J, Pojar S, Dos Santos Pereira JN, *et al*. Global genetic analyses reveal strong inter-ethnic variability in the loss of activity of the organic cation transporter OCT1. *Genome Med* 2015;7(1):56. doi:10.1186/s13073-015-0172-0, PMID:26157489.
- [298] Semiz S, Dujic T, Causevic A. Pharmacogenetics and personalized treatment of type 2 diabetes. *Biochem Med (Zagreb)* 2013;23(2):154–171. doi:10.11613/bm.2013.020, PMID:23894862.
- [299] Dujic T, Causevic A, Bego T, Malenica M, Veljica-Asimi Z, Pearson ER, *et al*. Organic cation transporter 1 variants and gastrointestinal side effects of metformin in patients with Type 2 diabetes. *Diabet Med* 2016;33(4):511–514. doi:10.1111/dme.13040, PMID:26605869.
- [300] Dujic T, Zhou K, Donnelly LA, Tavendale R, Palmer CN, Pearson ER. Association of Organic Cation Transporter 1 With Intolerance to Metformin in Type 2 Diabetes: A GoDARTS Study. *Diabetes* 2015;64(5):1786–1793. doi:10.2337/db14-1388, PMID:25510240.
- [301] Koshy M, Sethupathy S, Annamalai PT, Renju VC, Santha K. Association of oct1 gene polymorphism with glycemic status and serum metformin levels in type ii diabetes mellitus patients. *Int J Pharm Sci Res* 2013;4(5):1940–1945. doi:10.13040/IJPSR.0975-8232.4(5).1940-45.
- [302] Koepsell H, Lips K, Volk C. Polyspecific organic cation transporters: structure, function, physiological roles, and biopharmaceutical implications. *Pharm Res* 2007;24(7):1227–1251. doi:10.1007/s11095-007-9254-z, PMID:17473959.
- [303] Kang HJ, Song IS, Shin HJ, Kim WY, Lee CH, Shim JC, *et al*. Identification and functional characterization of genetic variants of human organic cation transporters in a Korean population. *Drug Metab Dispos* 2007;35(4):667–675. doi:10.1124/dmd.106.013581, PMID:17220237.
- [304] Leabman MK, Huang CC, Kawamoto M, Johns SJ, Stryke D, Ferrin TE, *et al*. Pharmacogenetics of Membrane Transporters Investigators. Polymorphisms in a human kidney xenobiotic transporter, OCT2, exhibit altered function. *Pharmacogenetics* 2002;12(5):395–405. doi:10.1097/00008571-200207000-00007, PMID:12142729.
- [305] Zolk O, Solbach TF, König J, Fromm MF. Functional characterization of the human organic cation transporter 2 variant p.270Ala>Ser. *Drug Metab Dispos* 2009;37(6):1312–1318. doi:10.1124/dmd.108.023762, PMID:19251820.
- [306] Shingaki T, Hume WE, Takashima T, Katayama Y, Okauchi T, Hayashinaka E, *et al*. Quantitative Evaluation of mMate1 Function Based on Minimally Invasive Measurement of Tissue Concentration Using PET with [(11)C]Metformin in Mouse. *Pharm Res* 2015;32(8):2538–2547. doi:10.1007/s11095-015-1642-1, PMID:25715695.

- [307] Chen Y, Teranishi K, Li S, Yee SW, Hesselton S, Stryke D, *et al*. Genetic variants in multidrug and toxic compound extrusion-1, hMATE1, alter transport function. *Pharmacogenomics J* 2009;9(2):127–136. doi:10.1038/tpj.2008.19, PMID:19172157.
- [308] Toyama K, Yonezawa A, Masuda S, Osawa R, Hosokawa M, Fujimoto S, *et al*. Loss of multidrug and toxin extrusion 1 (MATE1) is associated with metformin-induced lactic acidosis. *Br J Pharmacol* 2012; 166(3):1183–1191. doi:10.1111/j.1476-5381.2012.01853.x, PMID:2242910.
- [309] Morales-Rivera MI, Alemón-Medina R, Martínez-Hernández A, Gómez-Garduño J, Mirzaeicheshmeh E, Altamirano-Bustamante NF, *et al*. The L125F MATE1 variant enriched in populations of Amerindian origin is associated with increased plasma levels of metformin and lactate. *Biomed Pharmacother* 2021;142:112009. doi:10.1016/j.biopha.2021.112009, PMID:34388523.
- [310] Choi JH, Yee SW, Ramirez AH, Morrissey KM, Jang GH, Joski PJ, *et al*. A common 5'-UTR variant in MATE2-K is associated with poor response to metformin. *Clin Pharmacol Ther* 2011;90(5):674–684. doi:10.1038/clpt.2011.165, PMID:21956618.
- [311] Chung JY, Cho SK, Kim TH, Kim KH, Jang GH, Kim CO, *et al*. Functional characterization of MATE2-K genetic variants and their effects on metformin pharmacokinetics. *Pharmacogenet Genomics* 2013;23(7):365–373. doi:10.1097/FPC.0b013e3283622037, PMID:23652408.
- [312] Li Q, Li C, Li H, Zeng L, Kang Z, Mao Y, *et al*. STK11 rs2075604 Polymorphism Is Associated with Metformin Efficacy in Chinese Type 2 Diabetes Mellitus. *Int J Endocrinol* 2017;2017:3402808. doi:10.1155/2017/3402808, PMID:28775741.
- [313] Hu X-F, Zhang W-Y, Wen Q, Chen W-J, Wang Z-M, Chen J, *et al*. Fecal microbiota transplantation alleviates myocardial damage in myocarditis by restoring the microbiota composition. *Pharmacol Res* 2019;139:412–21. doi:10.1016/j.phrs.2018.11.042, PMID:30508676.
- [314] Wang H, Lu Y, Yan Y, Tian S, Zheng D, Leng D, *et al*. Promising Treatment for Type 2 Diabetes: Fecal Microbiota Transplantation Reverses Insulin Resistance and Impaired Islets. *Front Cell Infect Microbiol*. 2020;17 9:455. doi:10.3389/fcimb.2019.00455, PMID:32010641.
- [315] Broadfield LA, Saigal A, Szamosi JC, Hammill JA, Bezverbnaya K, Wang D, *et al*. Metformin-induced reductions in tumor growth involves modulation of the gut microbiome. *Mol Metab* 2022;61:101498. doi:10.1016/j.molmet.2022.101498, PMID:35452877.
- [316] Sachdeva M, Sra HK, Agarwal A, Chauhan A, Pradhan P, Singh M, *et al*. Effect of Probiotics on the Frequency of CD4+ T-Cells in HIV-Infected Children and Adolescents: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *J Trop Pediatr* 2022;68(2):fmac006. doi:10.1093/tropej/fmac006, PMID:35137236.
- [317] Xiang S, Ji JL, Li S, Cao XP, Xu W, Tan L, *et al*. Efficacy and Safety of Probiotics for the Treatment of Alzheimer's Disease, Mild Cognitive Impairment, and Parkinson's Disease: A Systematic Review and Meta-Analysis. *Front Aging Neurosci* 2022;14:730036. doi:10.3389/fnagi.2022.730036, PMID:35185522.